

Validation data for THP1-Dual™ KO-TLR8 Cells

<https://www.invivogen.com/thp1-dual-ko-trl8>

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Version 21B04-ED

THP1-Dual™ KO-TLR8 cells were generated from the THP1-Dual™ cell line through the verified stable knockout of the TLR8 gene (Figure 1). These cells feature two reporter genes allowing the simultaneous study of the NF-κB pathway by monitoring the activity of an inducible SEAP (secreted embryonic alkaline phosphatase)(Figure 2) and the IRF pathway by monitoring the activity of an inducible secreted Lucia luciferase (Figure 3).

Validation of TLR8 knockout

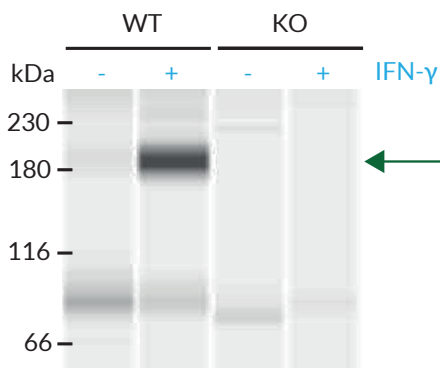


Figure 1: Validation of TLR8 KO. PMA-differentiated THP1-Dual™ (WT) and THP1-Dual™ KO-TLR8 (KO) cells were treated with 20 ng/ml interferon(IFN)-γ to induce the expression of TLR8. Lysates from non-treated (-) and treated (+) WT and KO cells were analyzed using an anti-human TLR8 antibody, followed by an HRP-conjugated anti-rabbit secondary antibody (WES assay). As expected, a band was detected in the IFN-γ treated WT cells only (green arrow). The predicted molecular weight of TLR8 is ~120 kDa.

Functional validation of TLR8 knockout (NF-κB response)

TLR8 recognizes microbial RNA degradation products, which ultimately leads to MyD88-dependent activation of NF-κB. As expected, the NF-κB response is abolished in THP1-Dual™ KO-TLR8 cells upon incubation with both synthetic ligands (R848 & CL075) and single-stranded (ss) RNA (ORN06 & ssRNA40) when compared to the THP1-Dual™ cells. No notable differences are observed for the other ligands tested.

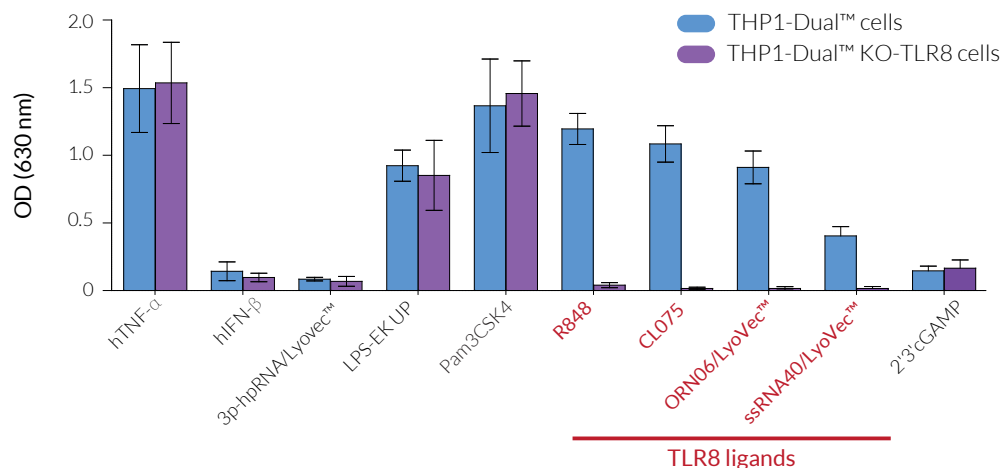


Figure 2: NF-κB responses in THP1-Dual™-derived cells. THP1-Dual™ and THP1-Dual™ KO-TLR8 cells were incubated with 0.3 ng/ml human (h)TNF-α (NF-κB-SEAP positive control), 1 x 10⁴ U/ml hIFN-β (IRF-Lucia positive control), 300 ng/ml 3p-hpRNA/LyoVec™ (RIG-I agonist), 1 μg/ml LPS-EK Ultrapure (UP; TLR4), 0.1 ng/ml Pam3CSK4 (TLR2 agonist), 10 μg/ml R848 (TLR7/8 agonist), 10 μg/ml CL075 (TLR7/8 agonist), 5 μg/ml ORN06/LyoVec™ (TLR8 agonist), 5 μg/ml ssRNA40/LyoVec™ (TLR8 agonist) and 10 μg/ml 2'3'-cGAMP (STING agonist). After overnight incubation, the activation of NF-κB was assessed by measuring the activity of SEAP in the supernatant using QUANTI-Blue™ Solution. Data are shown as optical density (OD) at 630 nm (mean ± SEM).

TECHNICAL SUPPORT

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Functional validation of TLR8 knockout (IRF response)

TLR8 has been shown to activate IRF-dependent signaling. THP1-Dual™ (parental cell line) displays a slight increase in the IRF-dependent secretion of Lucia luciferase upon activation with TLR8-specific agonists (in red). Importantly, this response is abolished in THP1-Dual™ KO-TLR8 cells. Additionally, the response of known ligands to signal in an IRF-dependent manner, such as 2'3'-cGAMP (STING agonist) and human interferon β (hIFN- β) was confirmed to be unchanged between THP1-Dual™ and THP1-Dual™ KO-TLR8.

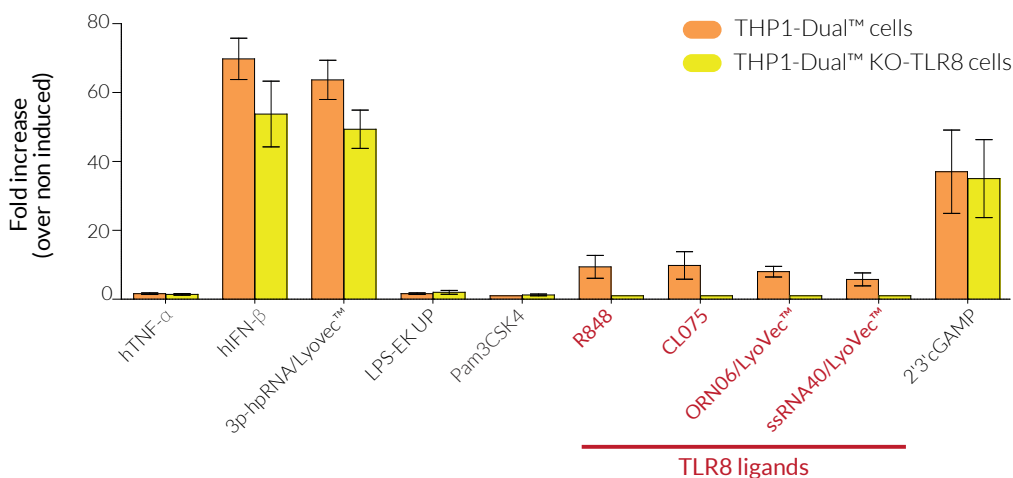


Figure 3: IRF responses in THP1-Dual™-derived cells. THP1-Dual™ and THP1-Dual™ KO-TLR8 cells were incubated with 0.3 ng/ml human (h)TNF- α (NF- κ B-SEAP positive control), 1×10^4 U/ml hIFN- β (IRF-Lucia positive control), 300 ng/ml 3p-hpRNA/LyoVec™ (RIG-I agonist), 1 μ g/ml LPS-EK Ultrapure (UP; TLR4), 0.1 ng/ml Pam3CSK4 (TLR2 agonist), 10 μ g/ml R848 (synthetic TLR7/8 agonist), 10 μ g/ml CL075 (synthetic TLR7/8 agonist), 5 μ g/ml ORN06/LyoVec™ ('natural' TLR8 agonist), 5 μ g/ml ssRNA40/LyoVec™ ('natural' TLR8 agonist) and 3 μ g/ml 2'3'-cGAMP (STING agonist). After overnight incubation, the IRF response was assessed by measuring the activity of Lucia luciferase in the supernatant using QUANTI-Luc™. Data are shown as a fold change (mean \pm SEM) over non-induced cells.

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