THP1-Blue™ KI-IP10 Cells

CXCL10 Knockin Lucia luciferase gene and NF-κB SEAP Reporter Monocytes

Catalog code: thpb-ip10kilc

https://www.invivogen.com/thp1-blue-ki-ip10

For research use only

Version 23E23-MM

PRODUCT INFORMATION

Contents and Storage

- **3-7 x 10⁶ of THP1-Blue™ KI-IP10 cells in a cryovial or shipping flask.** *IMPORTANT: If cells provided in a cryovial are not frozen upon arrival, contact InvivoGen immediately.*
- **1 ml of Blasticidin** (10 mg/ml). Store at 4 °C or at -20 °C.*
- **1 ml of Normocin™** (50 mg/ml), a formulation of three antibiotics active against mycoplasmas, bacteria and fungi. Store at -20°C.*

*The expiry date is specified on the product label.

- **1 tube of QUANTI-Luc™ 4 Reagent**, a Lucia luciferase detection reagent (sufficient to prepare 25 ml). Store at -20 °C. Avoid repeated freeze-thaw cycles. Note: *This product is photosensitive and should be protected from light.*
- **1 ml of QB reagent** and **1 ml of QB buffer** (sufficient to prepare 100 ml of **QUANTI-Blue™ Solution**, a SEAP detection reagent). Store QB reagent and QB buffer at -20 °C. QUANTI-Blue™ Solution is stable for 2 weeks at 4 °C and for 2 months at -20 °C.

Note: *Data sheets for all components are available on our website.*

Handling Frozen Cells Upon Arrival

Cells must be thawed immediately upon receipt and grown according to handling procedures (as described on the next page) to ensure the best cell viability and proper assay performance.

Note: **Avoid freezing cells upon receipt** *as it may result in irreversible damage to the cell line.*

Disclaimer: *We cannot guarantee cell viability if the cells are not thawed immediately upon receipt and grown according to handling procedures.*

IMPORTANT: For cells that arrive in a shipping flask please refer to the enclosed 'cell recovery procedure'.

Cell Line Stability

Cells will undergo genotypic changes over time resulting in reduced responsiveness in normal cell culture conditions. Genetic instability is a biological phenomenon that occurs in all stably transfected cells. Therefore, it is critical to prepare an adequate number of frozen stocks at early passages. THP1-Blue™ KI-IP10 cells should not be passaged more than 20 times to remain fully functional.

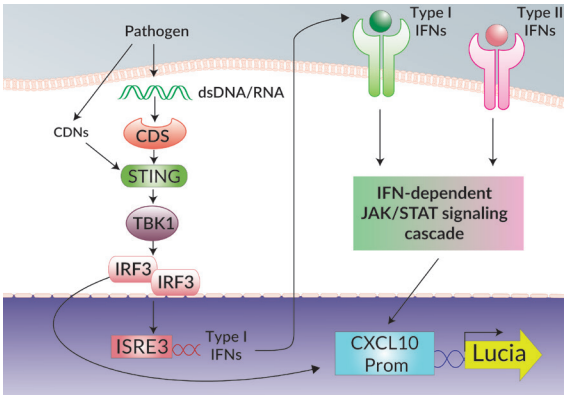
Quality Control

- Lucia luciferase gene knockin for IP10/CXCL10 was verified by PCR and DNA sequencing.
- THP1-Blue™ KI-IP10 cells have been stimulated with and respond to type I IFNs as well as various agonists of STING, Toll-like receptors (TLR) and RIG-I-like receptors (RLRs).
- The stability for 20 passages following thawing has been verified.
- THP1-Blue™ KI-IP10 cells are guaranteed mycoplasma-free.

BACKGROUND

Interferon-inducible protein-10 (IP10), more commonly known as CXCL10 (C-X-C motif ligand 10), belongs to a family of chemokines that includes CXCL9 and CXCL11. Canonically, CXCL10 is identified as an interferon stimulated gene (ISG) due to its induction by the JAK/STAT signaling cascades activated by type I and II IFNs[1]. Additionally, CXCL10 is strongly induced by the activation of the STING/TBK1/IRF3 signaling axis.

This cytosolic DNA-sensing pathway is activated by recognition of pathogen-derived nucleic acids and cyclic dinucleotides (CDNs) by cytosolic DNA sensors (CDS) and STING, respectively[2]. CXCL10 expression parallels the host's pro-inflammatory response to a number of pathogens including viruses such as respiratory syncytial virus, bacteria such as *Helicobacter pylori*, and parasites such as *Plasmodium falciparum*[3]. Therefore, CXCL10 expression has been suggested as a biomarker of disease severity. CXCL10 is a pleiotropic molecule that acts on both innate and adaptive immune cells. These diverse actions make it capable of exerting potent biological functions including apoptosis and the regulation of cell proliferation in infectious and inflammatory diseases. Interestingly, CXCL10 inhibits angiogenesis and therefore exerts additional important anti-tumor effects[3].



1. Muller, M. *et al.*, 2010. The chemokine receptor CXCR3 and its ligands CXCL9, CXCL10 and CXCL11 in neuroimmunity-a tale of conflict and conundrum. Neuropathol Appl Neurobiol. 36:368-387. 2. Motani, K. *et al.*, 2015. DNA-Mediated Cyclic GMP-AMP Synthase-Dependent and -Independent Regulation of Innate Immune Responses. J Immunol. 194:4914-4923. 3. Liu, M. *et al.*, 2011. CXCL10/IP-10 in infectious diseases pathogenesis and potential therapeutic implications. Cytokine Growth Factor Rev. 22:121-130.

CELL LINE DESCRIPTION

THP1-Blue™ KI-IP10 cells have been engineered from the human monocyte cell line THP1-Blue™ NF-κB to examine specifically the endogenous expression of CXCL10 in response to bacterial and viral infection, while preserving its genomic environment. In these cells, the CXCL10 open reading frame has been replaced with a Lucia luciferase reporter gene (from ATG to TGA), maintaining its expression under the control of the endogenous CXCL10 promoter. This allows the specific study of CXCL10 expression upon stimulation with agonists such as nucleic acids and cyclic dinucleotides (CDNs). The induction of endogenous CXCL10 expression can be easily quantified by assessing the expression of Lucia luciferase using the detection reagent QUANTI-Luc™ 4 Lucia/Gaussia.Furthermore, the activation of the NF-κB pathway, using an agonist such as tumor necrosis factor-α (TNF-α), is convenient for cross sample normalization by monitoring the activation of SEAP using the detection reagent QUANTI-Blue™ Solution.

THP1-Blue™ KI-IP10 cells are resistant to blasticidin.

TECHNICAL SUPPORT
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FAQ Any questions about our cell lines?
Visit our FAQ page.

InvivoGen
www.invivogen.com

SAFETY CONSIDERATIONS

Biosafety Level 1

HANDLING PROCEDURES

Required Cell Culture Medium

- **Growth Medium:** RPMI 1640, 10% (v/v) heat-inactivated fetal bovine serum (HI-FBS; 30 min at 56°C), 100 µg/ml Normocin™, Pen-Strep (100 U/ml-100 µg/ml)

Initial culture of all THP-1 derived cells must be performed in growth medium containing 20% heat-inactivated FBS.

Note: The use of Normocin™ together with Pen-Strep is required to keep the cells free of microbial contaminants. Contamination of this cell line may activate TLRs resulting in differentiation of the monocytes and activation of the reporter gene.

- **Freezing Medium:** 95% FBS, 5% DMSO
- **Test Medium:** RPMI 1640, 10% heat-inactivated FBS, Pen-Strep (100 U/ml-100 µg/ml).

Required Selective Antibiotics

Blasticidin

Initial Culture Procedure

The first propagation of cells should be for generating stocks for future use. This ensures the ongoing stability and performance of the cells.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid.
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol.

Note: All steps from this point should be carried out under aseptic conditions.

3. Transfer the cells to a tube containing 15 ml of pre-warmed growth medium (with 20% heat-inactivated FBS). **Do not add selective antibiotics until the cells have been passaged twice.**
4. Centrifuge the tube at 150 x g (RCF) for 10 mins.
5. Remove the supernatant containing the cryoprotective agent and resuspend the cells with 1 ml of growth medium (with 20% heat-inactivated FBS).
6. Transfer the cells (1 ml) to a 25 cm^2 tissue culture flask containing 5 ml of growth medium (with 20% heat-inactivated FBS).
7. Place the culture at 37°C and 5% CO_2.

Frozen Stock Preparation

1. Resuspend cells to a density of 5-7 x 10^6 cells/ml in freshly prepared freezing medium.
2. Dispense 1 ml aliquots of the cell suspension into cryogenic vials.
3. Place the vials in a freezing container and store at -80°C overnight.
4. Transfer the vials to liquid nitrogen for long term storage.

Note: If properly stored, cells should remain stable for years.

Cell Maintenance

1. After cells have recovered (after at least two passages), subculture the cells in growth medium (with 10% heat-inactivated FBS). To maintain selection pressure, add 10 µg/ml of Blasticidin to the growth medium every other passage.
2. Passage the cells every 3 days by inoculating 5 x 10^5 cells/ml. Do not allow the cell concentration to exceed 2 x 10^6 cells/ml.

Cell Handling Recommendations

- Use THP1-Blue™ KI-IP10 cells with less than 20 passages.

- Handling of cells should be as short as possible to prevent any damage resulting from a prolonged stay at room temperature without 5% CO_2

REPORTER ASSAYS

Cell Suspension Preparation

1. Centrifuge cells at 150 x g (RCF) for 10 mins or 300 x g (RCF) for 5 mins.
2. Remove supernatant and resuspend THP1-Blue™ KI-IP10 cells to 5 x 10^5 cells/ml in freshly prepared test medium.

Note: It is recommended to perform the assay with test medium, which does not contain Normocin™ or Blasticidin.

Detection of IP10 (CXCL10) induction

Below is a protocol for end-point readings using a luminometer with an injector, however, this protocol can be adapted for use with kinetic measurements or a luminometer with a manual set-up.

1. Add 20 µl of sample per well of a flat-bottom 96-well plate including 2'3' cGAMP as a positive control and endotoxin free water as a negative control (use new tips for each well to avoid cross-contamination).
2. Add 180 µl of the cell suspension (~100,000 cells) per well.
3. Incubate the plate for 18-24 h at 37°C and 5% CO_2.
4. Prepare QUANTI-Luc™ 4 Reagent working solution following the instructions on the enclosed data sheet.
5. Setup the luminometer with the following parameters: 50 µl of injection, end-point measurement with a 4 second start time and 0.1 second reading time.
6. Add 10 µl of the THP1-Blue™ KI-IP10 cells supernatant into a 96-well white (opaque) or black plate, or a luminometer tube.
7. Prime the injector with QUANTI-Luc™ 4 Reagent working solution and proceed with the measurement.

Detection of NF-κB stimulation

1. Add 20 µl of sample per well of a flat-bottom 96-well plate, including a positive control (such as TNF-α) and endotoxin free water as a negative control (use new tips for each well to avoid cross-contamination).
2. Add 180 µl of the cell suspension (~100,000 cells) per well.
3. Incubate the plate for 18-24 h at 37°C and 5% CO_2.
4. Prepare QUANTI-Blue™ Solution following the instructions on the enclosed data sheet.
5. Add 180 µl of reconstituted QUANTI-Blue™ Solution per well of a flat-bottom 96-well plate.
6. Add 20 µl of the THP1-Blue™ KI-IP10 cells supernatant.
7. Incubate the plate at 37°C for 1-6 h.
8. Determine SEAP levels using a spectrophotometer at 620-655 nm.

USE RESTRICTIONS

These cells are distributed for research purposes only.

This product is covered by a Limited Use License. By use of this product the buyer agrees to the terms and conditions of all applicable Limited Use Label Licenses. For non-research use, such as screening, quality control or clinical development, contact info@invivogen.com.

RELATED PRODUCTS

Product	Description	Cat. Code
2'3'-cGAMP	Cyclic [G(2',5')pA(3',5')p]	tlrl-nacga23
Blasticidin	Selection antibiotic	ant-bl-1
Normocin™	Antimicrobial antibiotic	ant-nr-1
QUANTI-Blue™ Solution	SEAP detection reagent	rep-qbs1
QUANTI-Luc™ 4 Lucia/Gaussia	Luminescence detection kit	rep-qlc4lg1
Recombinant hTNF-α	Recombinant cytokine	rcyc-htnfa

TECHNICAL SUPPORT
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QUANTI-Blue™ Solution

Medium for detection and quantification of alkaline phosphatase in standard and HTS assays

Catalog code: rep-qbs, rep-qbs2, rep-qbs3

https://www.invivogen.com/quanti-blue

For research use only

Version 23C09-MM

PRODUCT INFORMATION

Contents: QUANTI-Blue™ Solution is available in three pack sizes

- **rep-qbs:** 5 x 1 ml of QB reagent and 5 x 1 ml QB buffer, sufficient to prepare QUANTI-Blue™ Solution for **25 x 96-well plates** (500 ml using the standard procedure) or **20 x 1536-well plates** (85 ml using the HTS screening procedure).
- **rep-qbs2:** 10 x 1 ml of QB reagent and 10 x 1 ml QB buffer, sufficient to prepare QUANTI-Blue™ Solution for **50 x 96-well plates** (1 L using the standard procedure) or **40 x 1536-well plates** (170 ml using the HTS screening procedure).
- **rep-qbs3:** 1 x 20 ml bottle of QB reagent and 1 x 20 ml bottle of QB buffer, sufficient to prepare QUANTI-Blue™ Solution for **100 x 96-well plates** (2 L using the standard procedure) or **80 x 1536-well plates** (340 ml using the HTS screening procedure).

Required Material (not provided)

- Sterile water
- Sterile screw cap tube, glass bottle or flask

Storage and stability

- Product is shipped at room temperature. Upon receipt, store QB reagent and QB buffer at -20 °C. Product is stable for 1 year at -20 °C when properly stored.
- The 20 ml bottles of QB reagent and QB buffer are designed for single use. If required, individual aliquots of QB reagent and QB buffer can be prepared upon receipt or following a single freeze-thaw cycle. Store aliquots at -20°C. **Avoid repeated freeze-thaw cycles**.

Note: During storage, a precipitate may form in the 20 ml bottle of QB reagent and QB buffer. If this occurs, heat the product at 37°C for 30 seconds and vortex until the precipitate disappears. The formation of a precipitate does not affect the activity of the product.

- Reconstituted QUANTI-Blue™ Solution is stable for 2 weeks at 2-8 °C and for 2 months at -20°C. Protect from light.

Quality Control

Each lot is thoroughly tested to ensure the absence of lot-to-lot variation.

- Physicochemical characterization (pH, appearance).
- Functional assays using alkaline phosphatase or SEAP-expressing reporter cells.

DESCRIPTION

QUANTI-Blue™ is a colorimetric enzyme assay developed to determine any alkaline phosphatase activity (AP) in a biological sample, such as supernatants of cell cultures. QUANTI-Blue™ Solution changes from pink to a purple-blue color in the presence of AP. Secreted embryonic alkaline phosphatase (SEAP) is a widely used reporter gene. It is a truncated form of placental alkaline phosphatase, a glycosylphosphatidylinositol (GPI)-anchored protein. SEAP is secreted into the cell culture supernatant and therefore offers many advantages over intracellular reporters.

QUANTI-Blue™ is highly sensitive for quantitative measurement. It has a higher saturation threshold than with pNPP (p-nitrophenyl phosphate) resulting in more significant differences between no, low or high AP activity. Another advantage of QUANTI-Blue™ is that it can determine secreted AP activity without disturbing cells, thus allowing the repeated sampling of cell cultures for kinetic studies.

METHODS

QUANTI-Blue™ Solution has been optimized for use in 96-well plates (standard procedure) and in 1536-well plates (high throughput screening procedure).

A. Standard procedure

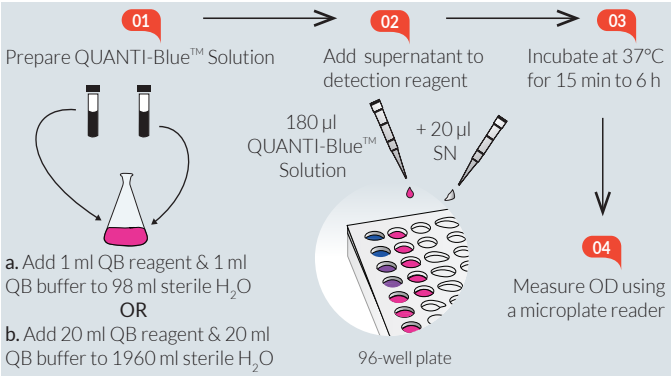

Figure 1. Standard procedure using QUANTI-Blue™ Solution.

The following protocol refers to the use of 96-well plates. Ensure QB reagent and QB buffer are completely thawed before use.

*Note: For fast thawing, QB reagent and QB buffer can be placed at 37 °C for 2 minutes. Ensure heating at 37 °C does **not** exceed 5 minutes.*

1. In a sterile bottle or flask, prepare QUANTI-Blue™ Solution by adding:
   a. **1 ml** of QB reagent and **1 ml** of QB buffer to **98 ml** of sterile water.
   b. **20 ml** of QB reagent and **20 ml** of QB buffer to **1960 ml** of sterile water.
2. Mix by vortexing and incubate at room temperature for 10 min before use.
3. Use QUANTI-Blue™ Solution immediately or store at 2-8 °C or -20°C.
4. Dispense 180 µl of QUANTI-Blue™ Solution per well into a flat-bottom 96-well plate.
5. Add 20 µl of the sample (supernatant of SEAP-expressing cells) or negative control (cell culture medium).
6. Incubate at 37 °C for 15 min to 6 h.
7. Measure optical density (OD) at 620-655 nm using a microplate reader.

Note: If the negative control turns purple/blue, it means the fetal bovine serum (FBS) contains alkaline phosphatase. We recommend heating FBS at 56 °C for 30 min to inactivate the alkaline phosphatase activity.

For different cell culture plate formats, please refer to the table below:

	96-well plate	24-well plate	12-well plate
QUANTI-Blue™	180 µl	450 µl	900 µl
Supernatant	20 µl	50 µl	100 µl

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B. High Throughput Screening (HTS) procedure

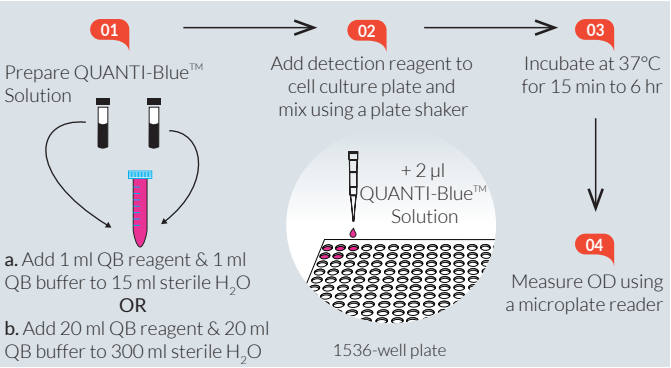

Figure 2. High throughput screening procedure using QUANTI-Blue™ Solution.

This procedure has been optimized for use in HTS screening procedures in 1536-well plates. QUANTI-Blue™ Solution is added directly to the cell suspension to reduce liquid handling.
Ensure QB reagent and QB buffer are completely thawed before use.
Note: For fast thawing, QB reagent and QB buffer can be placed at 37 °C for 2 minutes. Ensure heating at 37 °C does ***not*** *exceed 5 minutes.*

1. Dispense cell suspension and test compounds into a 1536-well plate in a volume that does not exceed **5 µl** per well. Incubate cells with test compounds for the desired period of time.
2. Prepare QUANTI-Blue™ Solution by adding:
   a. **1 ml** of QB reagent and **1 ml** of QB buffer to **15 ml** of sterile water in a sterile 50 ml screw cap tube.
   b. **20 ml** of QB reagent and **20 ml** of QB buffer to **300 ml** of sterile water in a sterile glass bottle or flask.
3. Mix well by vortexing and incubate at room temperature for 10 minutes before use.
4. Use QUANTI-Blue™ Solution immediately or store at 2-8 °C or -20 °C.
5. Dispense **2 µl of QUANTI-Blue™ Solution** to the wells containing ≤ 5 µl of cell culture in a 1536-well plate.
6. Mix using a plate shaker.
7. Incubate at 37 °C for 15 min to 6 h.
8. Measure OD at 620-655 nm.

Note: If the negative control turns purple/blue, it means the fetal bovine serum (FBS) contains alkaline phosphatase. We recommend heating FBS at 56 °C for 30 min to inactivate the alkaline phosphatase activity.

RELATED PRODUCTS

Product	Catalog Code
pNiFty2-SEAP (Zeo®)	pnifty2-seap
pSELECT-zeo-SEAP	psetz-seap
HEK-Blue™ Detection	hb-det2
Recombinant SEAP Protein	rec-hseap
Reporter cells	
HEK-Blue™ hTLR2	hkb-htlr2
HEK-Blue™ hTLR4	hkb-htlr4
RAW-Blue™ Cells	raw-sp
THP1-Blue™ NF-κB Cells	thp-nfkb
THP1-Blue™ ISG Cells	thp-isg

For a complete list of InvivoGen's Reporter Cell Lines visit
https://www.invivogen.com/reporter-cells

QUANTI-Luc™ 4 Reagent

A coelenterazine-based luminescence assay reagent

https://www.invivogen.com/quanti-luc

For research use only

Version 23C24-AK

PRODUCT INFORMATION

Contents

- 1 tube of **QUANTI-Luc™ 4 Reagent (20X)**

One tube of QUANTI-Luc™ 4 Reagent is sufficient for 5 x 96-well plates (25 ml standard Flash/end-point detection).

Note: This sample cannot be sold separately from the QUANTI-Luc™ 4 Lucia/Gaussia kit.

QUANTI-Luc™ 4 Lucia/Gaussia comprises two liquid components:

- QUANTI-Luc™ 4 Reagent 20X (coelenterazine substrate)
- QUANTI-Luc™ 4 Stabilizer 25X (optimized Glow assay reagent)

Find more information at https://www.invivogen.com/quanti-luc.

Storage and Stability

– Store QUANTI-Luc™ 4 Reagent at -20°C for up to 12 months.
– After preparation, the working solution is stable for 48 hours at 4°C and 1 month at -20°C. Prepare aliquots to avoid repeated freeze-thaw cycles.
Note: This product is photosensitive and should be protected from light.

Quality Control

Each lot is thoroughly tested to ensure the absence of lot-to-lot variation.
– Physicochemical characterization (pH, appearance).
– Functional assays using recombinant Lucia protein or reporter cells.

DESCRIPTION

QUANTI-Luc™ 4 Reagent is a component of the QUANTI-Luc™ 4 Lucia/Gaussia kit. It contains the coelenterazine substrate for the detection of secreted Lucia or Gaussia activity in live-cell supernatants, and of intracellular Renilla after cell lysis. The light signal produced correlates to the amount of luciferase protein expressed. It is quantified using a luminometer and expressed as relative light units (RLUs).

METHODS

Preparation of QUANTI-Luc™ 4 Reagent working solution (1X):

1. Dilute the total volume of the 20X tube (1.25 ml) of Reagent into 23.75 ml of sterile water to obtain 25 ml of working solution.
2. Vortex **very briefly** (a few seconds).
3. Use the working solution immediately or store until required for use. QUANTI-Luc™ 4 Reagent working solution can be stored for 48 hours at 4°C or 1 month at -20°C.

Flash detection of luciferase activity from cell culture medium:

To obtain **end-point readings** using a luminometer **with an injector**.
1. Set the luminometer with the following parameters: 50 µl of injection, end-point measurement with a 4 second start time and 0.1 second reading time.
2. Pipet 10-20 µl of sample per well into a 96-well white (opaque) or black plate, or a luminometer tube.
3. Prime the injector with QUANTI-Luc™ 4 Reagent 1X and proceed **immediately** with the measurement.

To obtain **end-point readings** using a luminometer **without injectors**.
1. Set the luminometer with a 0.1 second reading time.
2. Pipet 10-20 µl of sample per well into a 96-well white (opaque) or black plate, or a luminometer tube.
3. Add 50 µl of QUANTI-Luc™ 4 Reagent 1X to each well or tube.
4. Gently tap the plate several times to mix (do **not** vortex).
5. Proceed **immediately** with the measurement.

RELATED PRODUCTS

Product	Cat. Code
QUANTI-Luc™ 4 Lucia/Gaussia Kit	
500 tests	rep-qlc4lg1
2 x 500 tests	rep-qlc4lg2
5 x 500 tests	rep-qlc4lg5

TECHNICAL SUPPORT
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