

pFUSE2-CLIg-rk2

Plasmid featuring the constant region of the rabbit immunoglobulin kappa 2 light chain

Catalog # pfuse2-rlk2

For research use only

Version 20J02-MM

PRODUCT INFORMATION

Content:

- 20 µg of pFUSE2-CLIg-rk2 plasmid provided as lyophilized DNA.
- 2 x 1 ml blasticidin at 10 mg/ml

Storage and Stability:

- Product is shipped at room temperature.
- Lyophilized DNA should be stored at -20°C and is stable 3 months.
- Resuspended DNA should be stored at -20°C and is stable up to 1 year.

Quality control:

- Plasmid construct has been confirmed by restriction analysis and sequencing.
- Plasmid DNA was purified by ion exchange chromatography.

Materials required for antibody generation & isotype switching

- pFUSE2-CLIg plasmid that features the constant region of the kappa or lambda light chains. pFUSE2-CLIg plasmids are selectable with blasticidin.
- pFUSE-CHIg plasmid for the constant region of the heavy chain, this plasmid is selectable with Zeocin™ (sold separately, see RELATED PRODUCTS).

GENERAL PRODUCT USE

pFUSE-CLIg and pFUSE-CHIg plasmids are designed to change a monoclonal antibody from one isotype to another, therefore, enabling the generation of antibodies with the same antigen affinity but with different effector functions (increased or reduced ADCC and CDC). Furthermore, they can be used to produce entire IgG antibodies from Fab or scFv fragments.

pFUSE-CHIg and pFUSE2-CLIg express the constant regions of the heavy (CH) and light (CL) chains, respectively. They contain a multiple cloning site (MCS) upstream of these constant regions to enable the cloning of the variable (VH and VL) regions of a given antibody. Transfection of mammalian cell lines with the recombinant pFUSE-CHIg and pFUSE2-CLIg pair allows to generate an IgG antibody that can be purified from the supernatant using the appropriate Protein A, Protein G or Protein L affinity chromatography.

Features of pFUSE2-CLIg and pFUSE-CHIg plasmids

- **hEF1-HTLV prom** is a composite promoter comprising the Elongation Factor-1α (EF-1α) core promoter¹ and the R segment and part of the U5 sequence (R-U5') of the Human T-Cell Leukemia Virus (HTLV) Type 1 Long Terminal Repeat². The EF-1α promoter exhibits a strong activity and yields long lasting expression of a transgene *in vivo*. The R-U5' has been coupled to the EF-1α core promoter to enhance stability of RNA.
- **MCS:** The multiple cloning site contains several restriction sites that are compatible with many other enzymes, thus facilitating cloning.
- **SV40 pAn:** The Simian Virus 40 late polyadenylation signal enables efficient cleavage and polyadenylation reactions resulting in high levels of steady-state mRNA³.
- **ori:** A minimal *E. coli* origin of replication to limit vector size, but with the same activity as the longer Ori.
- **CMV enh / hFerL prom:** This composite promoter combines the human cytomegalovirus immediate-early gene 1 enhancer and the core promoter of the human ferritin light chain gene. This ubiquitous promoter drives the expression of the blasticidin-resistance gene in mammalian cells.
- **EM2KC** is a bacterial promoter that enables the constitutive expression of the antibiotic resistance gene in *E. coli*. EM2KC is located within an intron and is spliced out in mammalian cells.
- **βGlo pAn:** The human beta-globin 3'UTR and polyadenylation sequence allows efficient arrest of the transgene transcription⁴.

pFUSE2-CLIg-rk2 specific features

- **Rabbit IgKC2 (Ig Kappa 2 Light constant domain):** When cloning your VL region of choice in the MCS, care must be taken to preserve the integrity of the kappa 2 light chain constant region and reading frame.
- **Bsr (blasticidin resistance gene):** Resistance to blasticidin is conferred by the bsr gene from *Bacillus cereus*. The same resistance gene confers selection in both mammalian cells and *E. coli*.

References:

1. Kim DW. *et al.* 1990. Use of the human elongation factor 1 alpha promoter as a versatile and efficient expression system. 91(2):217-23. 2. Takebe Y. *et al.* 1988. SR alpha promoter: an efficient and versatile mammalian cDNA expression system composed of the simian virus 40 early promoter and the R-U5 segment of human T-cell leukemia virus type 1 long terminal repeat. Mol Cell Biol. 8(1):466-72. 3. Carswell S. & Alwine JC. 1989. Efficiency of utilization of the simian virus 40 late polyadenylation site: effects of upstream sequences. Mol Cell Biol. 9(10):4248-58. 4. Yu J. & Russell JE. 2001. Structural and functional analysis of an mRNP complex that mediates the high stability of human beta-globin mRNA. Mol Cell Biol. 21(17):5879-88.

TECHNICAL SUPPORT

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PROTOCOL

Obtaining VH and VL sequences

To obtain the cDNA sequence of the VH and VL regions from an antibody producing hybridoma, total RNA or mRNA is extracted and reverse transcribed to cDNA. PCR is performed with 5' degenerate primers to anneal to the unknown VH and VL regions and the 3' primers designed to anneal to the "known" CH and CL regions. Alternatively 5' RACE can be used. The resulting amplicons must be sequenced.

Plasmid resuspension

Quickly spin the tube containing the lyophilized plasmid to pellet the DNA. To obtain a plasmid solution at 1 µg/µl, resuspend the DNA in 20 µl of sterile H₂O. Store resuspended plasmid at -20°C.

Cloning into pFUSE-CHlg and pFUSE2-CLlg

Once the VH and VL sequence are known, inserts for cloning into the plasmids can be generated. In pFUSE2-CLlg-rk2, the constant region of the rabbit kappa 2 light chain is preceded by a multiple cloning site containing seven unique restriction sites: AgeI, EcoRI, EcoRV, XhoI, NcoI, Acc65I and BamHI. We recommend using the AgeI restriction site for insertion of the 5' end of the variable region including the native signal sequence. If the immunoglobulin signal sequence is unknown, pFUSE2ss and pFUSE2ess plasmids containing a signal sequence should be used. In pFUSE2-CLlg-rk2, use BamHI as the 3' cloning site for the VL in order to preserve the immunoglobulin kappa 2 constant amino acid sequence.

Notes:

- When generating the insert for VH, a XhoI restriction site must be introduced at the 3' end (for pFUSE-CHlg plasmid featuring the rabbit IgG (allele 3/2) heavy chain; see RELATED PRODUCTS). There is a choice of restriction sites at the 5' end.

- The 5' end of the variable region should encompass the native ATG initiation codon and the region immediately after which corresponds to the signal sequence. For proper initiation of translation, make sure that your insert contains a Kozak translation initiation sequence upstream of the ATG initiation codon such as (G/A)NNATG.

Antibody production

Cotransfect mammalian cells, such as 293 and CHO cells, with the recombinant plasmids pFUSE2-CLlg encoding the light chain and pFUSE-CHlg encoding the heavy chain. Antibody production depends greatly on the ratio of heavy chain and light chain expression. Typically, pFUSE-CHlg to pFUSE2-CLlg ratio of 2:3 is used to cotransfect mammalian cells. Since both plasmids share the same plasmid backbone, the appropriate heavy chain to light chain ratio can be easily determined by varying the quantities of plasmids.
OR

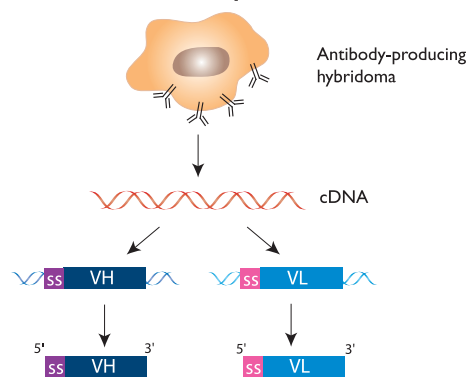
Transfect cells using a transfection agent, such as LyoVec™, with the plasmid coding for light chain and select the best clone. Following selection of the best clone, the plasmid coding for the heavy chain clone can be transfected into this clone.

Use blasticidin and Zeocin™ to select pFUSE2-CLlg and pFUSE-CHlg respectively.

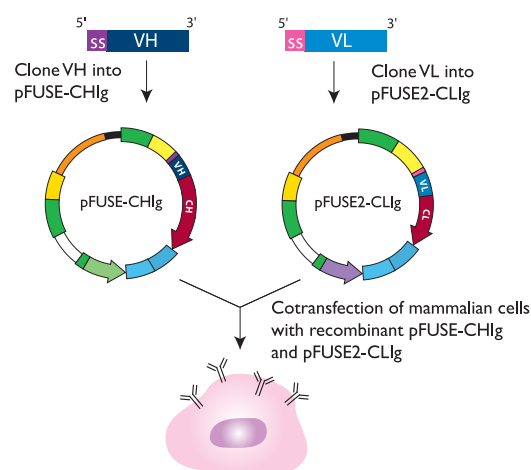
Antibody production can be analyzed by different techniques including SDS-PAGE, flow cytometry, ELISA, or a bioactivity assay.

Antibody generation using pFUSE-CHlg & pFUSE2-CLlg

1- Obtention of VH and VL sequences



2- Cloning into pFUSE-CHlg and pFUSE2-CLlg



Antibody purification

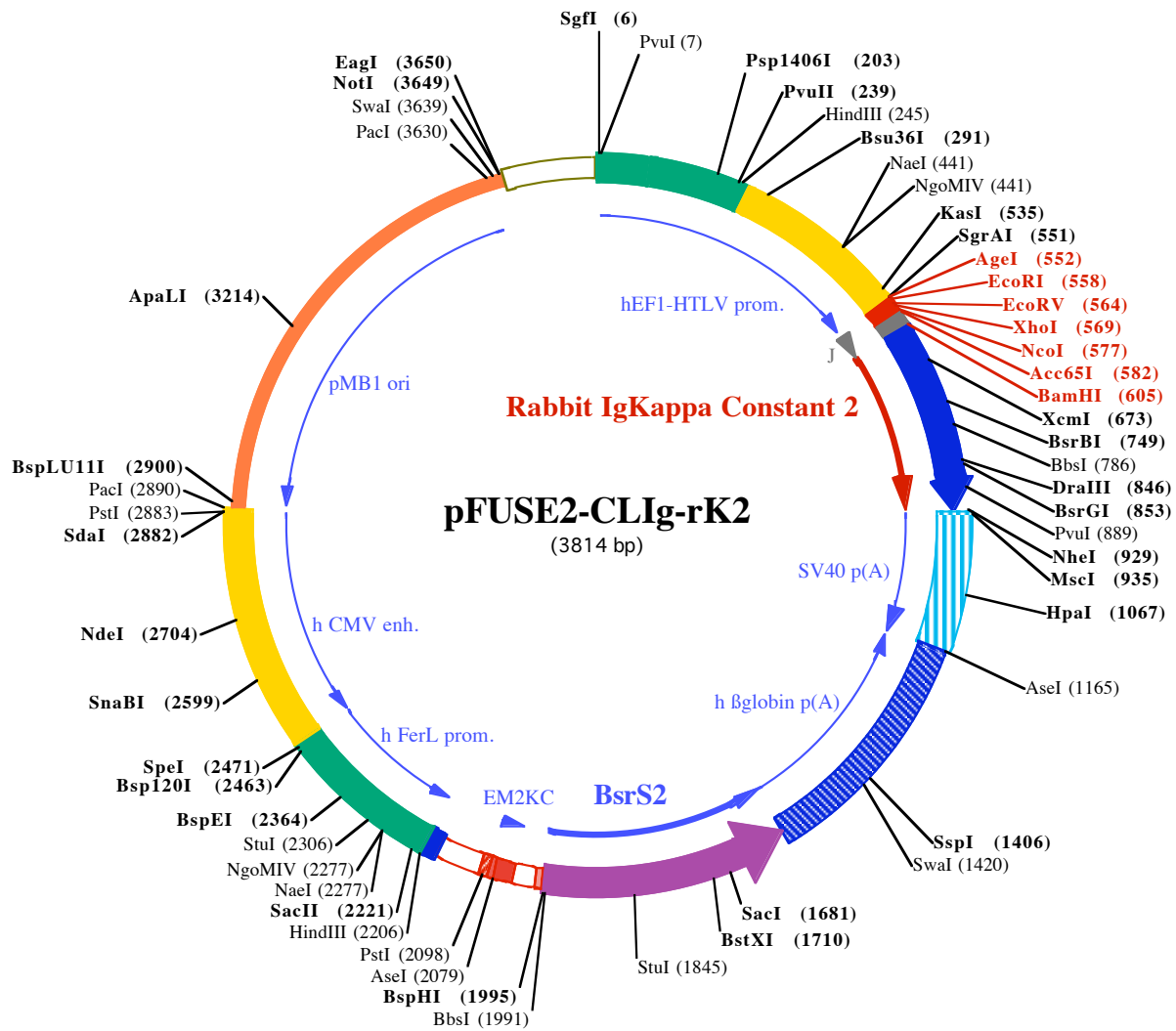
The resulting IgG antibody can be purified from the supernatant using the appropriate Protein A, Protein G or Protein L affinity chromatography.

RELATED PRODUCTS

Product	Catalog Code
pFUSE-CHlg-rG LyoVec™	pfuse-rchg lyec-12
Protein L / Agarose	gel-protl-2
Protein G / Agarose	gel-agg-5
Blasticidin	ant-bl-1

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100

PvuI (7)
SgfI (6)
1 GGATCTGGATCGCTCCGGTGCCCGTCAGTGGCAGAGCGCACATCGCCACAGTCCCGGAGAAGTTGGGGGAGGGGTCGGCAATTGAACGGGTGCCTA
101 GAGAAGGTGGCGCGGGTAAACTGGAAAGTGATGTCGTGTAAGTGGTCCGCCTTTTTCCGAGGGTGGGGGAGAACCCTATATAAGTGCAGTAGTCGCC

HindIII (245)
Psp1406I (203) PvuII (239) Bsu36I (291)
201 GTGAACGTTCTTTTTTCGCAACGGGTTTCCGCCAGAACACAGCTGAAGCTTCGAGGGGCTCGCATCTCTCTTACCGCCGCCGCCCTACCTGAGGCC
301 GCCATCCACGCCGGTTGAGTCGCGTTCTGCCGCTCCCGCCTGTGGTGCCTCTGAAGTGCCTCCGCCGTCTAGGTAAGTTTAAAGCTCAGGTCGAGACC

NgoMIV (441)
NaeI (441)
401 GGGCCTTTGTCCGGCGCTCCCTTGGAGCCTACCTAGACTCAGCCGGCTCTCCACGCTTTGCTGACCCTGCTTCTCAACTCTACGTCTTTGTTTCGTTT

EcoRI (558)
AgeI (552) XhoI (569) Acc65I (582)
KasI (535) SgrAI (551) EcoRV (564) NcoI (577)
501 TCTGTTCTGCGCCGTTACAGATCCAAGCTGTGACCGCGCCCTACCTGAGATCACCGTGAATTCGATATCTCGAGCCACATGGTACCAAGGTGGAATC
1 E F D I S S T M G T K V E I

BamHI (605) XcmI (673)
601 AAACGGGATCCAGTTGCGCCTTCTGTCCTCTTCCACCATCTAAGGAGGAGCTGACAAGTGAACAGCCACCATCGTGTGCGTGGCGAATAAATTCT
15 K R D P V A P S V L L F P P S K E E L T T G T A T I V C V A N K F

BsrBI (749) BbsI (786)
701 ATCCAGTGACATCACCGTCACCTGGAAGGTGGATGGCACCACCAACAGCCAGCGCATCGAGAACAGTAAAACACCGCAGAGCCCCGAAGACAATACCTA
32 Y P S D I T V T W K V D G T T Q Q S G I E N S K T P Q S P E D N T Y

BsrGI (853) PvuI (889)
801 CAGCCTGAGCAGCACTCTGCACTGACCAGCGCACAGTACAACAGCCACAGCGTGTACACCTGCGAGGTGGTCCAAGGCTCAGCCTCACCGATCGTCCAG
65 S L S S T L S L T S A Q Y N S H S V Y T C E V V Q G S A S P I V Q

MscI (935) NheI (929)
901 AGCTTCAATAGGGGTGACTGCTAGAGCGAGCTAGCTGGCCAGACATGATAAGATACATTGATGAGTTTGGACAAACCACAAGTGAATGCAGTAAAAAA
99 S F N R G D C •

HpaI (1067)
1001 ATGCTTTATTTGTGAAATTTGTGATGCTATTGCTTTATTTGTAACCATTATAAGCTGCAATAAACAAGTTAAACAACAATTGCATTCAATTTTATG

AseI (1165)
1098 TTTCAGTTTCAGGGGAGGTGTGGGAGGTTTTTAAAGCAAGTAAACCTCTACAATGTGGTATGGAATTAATTCTAAAATACAGCATAGCAAACTT
1197 TAACCTCAAATCAAGCCTCTACTTGAATCCTTTTCTGAGGGATGAATAAGGCATAGGCATCAGGGGCTGTTGCCAATGTGCATTAGCTGTTTGCAGCCT
1297 CACCTTCTTTCATGGAGTTAAGATATAGTGTATTTTCCAAGGTTTGAAGTACTCTTCATTCTTTATGTTTTAAATGCACTGACCTCCACATTCCC

SspI (1406) SmaI (1420)
1397 TTTTTAGTAAATATTCAGAAATAATTTAAATACATCATTGCAATGAAATAAATGTTTTTATTAGGCAGAATCCAGATGCTCAAGGCCCTTCATAATA
1497 TCCCCAGTTTAGTAGTTGGACTTAGGGAACAAAGGAACCTTTAATAGAAATTGGACAGCAAGAAAGCGAGCTTCTAGCTTTAGTTCTCTGGTGTACTTGA
141 • N R T Y K L

SacI (1681)
1597 GGGGATGAGTTCCTCAATGGTGGTTTTGACCAGCTTCCATTCTCAATGAGCACAAGCAGTCAGGAGCATAGTCAGAGATGAGCTCTCTGCACAT
134 P I L E E I T T K V L K G N M E I L V F C D P A Y D S I L E R C M

BstXI (1710)
1697 GCCACAGGGGCTGACCACCTGATGGATCTGTCCACCTCATCAGAGTAGGGGTGCCTGACAGCCACAATGGTGTCAAAGTCTTCTGCCCGTTGCTCACA
101 G C P S V V R I S R D V E D S Y P H R V A V I T D F D K Q G N S V

StuI (1845)
1797 GCAGACCAATGGCAATGGCTTCAGCACAGACAGTACCTGCCAATGTAGGCTCAATGTGGACAGCAGAGATGATCTCCAGTCTTGGTCTGATGG
67 A S G I A I A E A C V T V R G I Y A E I H V A S I I E G T K T R I A

BspHI (1991)
1897 CCGCCCCGACATGGTCTTGTGCTCCTCATAGAGCATGGTGTCTTCTCAGTGGCGACCTCCACCAGCTCCAGATCCTGCTGAGAGATGTTGAAGGCTCT
34 A G V H H K N D E Y L M T I K E T A V E V L E L D Q Q S I N F T K

AseI (2079)
1997 CATGATGGCTCTCctgtcaggagaggaagagaagaaggttagtacaattgCTATAGTGAGTTGTATTATACTATGCTTATGATTAATTGTCAAAC
1 M

PstI (2098)
2095 AGGGCTGCAgggttcagtgtccacttttctgactgccccatctctgcccaccctttccaggcatagacagtcagtacttacCAAACCTCACAGGA

HindIII (2206) SacII (2221) NgoMIV (2277) NaeI (2277)
2195 GGGAGAAGGCAGAAGCTTGAGACAGACCCGCGGACCGCCGAAGTGCAGGGGACGTGGCTAGGGCGGCTTTTTATGTTGCGCCGCCCTCGGAGGCA

StuI (2306) BspEI (2364)
2295 GGGCGCTCGGGGAGGCTAGCGGCAATCTGCGTGGCAGGAGCGGGCCGAAGGCCGTGCCTGACCAATCCGGAGCACATAGGAGTCTCAGCCCCCG

2395 CCCAAAGCAAGGGGAAGTCACGCGCTGTAGCGCCAGCGTGTGTGAAATGGGGGCTTGGGGGGTGGGGCCCTGACTAGTCAAACAAACTCCATT
Bsp120I (2463) SpeI (2471)
2495 GACGTCAATGGGGTGGAGACTTGAAATCCCCGTGAGTCAAACCGCTATCCACGCCATTGATGTACTGCCAAAACCGCATCATCATGGTAATAGCGATG

2595 ACTAATACGTAGATGTACTGCCAAGTAGAAAAGTCCATAAGGTCATGTACTGGGCATAATGCCAGGCGGGCCATTTACCGTCATTGACGTCAATAGGGG
SnaBI (2599)

2695 GCGTACTTGGCATATGATACACTTGATGTACTGCCAAGTGGGCAGTTTACCGTAAATACTCCACCCATTGACGTCAATGGAAAGTCCCTATTGGCGTTAC
NdeI (2704)

2795 TATGGGAACATACGTCATTATTGACGTCAATGGGCGGGGTCGTTGGGCGGTGAGCCAGGCGGGCCATTTACCGTAAGTTATGTAACGCGCTGCAGGTTAA
PstI (2883) SdaI (2882) PacI (2890)

2895 TTAAGAACATGTGAGCAAAGGCCAGCAAAGGCCAGGAACCGTAAAAAGGCCGCTTGTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCA
BspLU11I (2900)

2995 CAAAAATCGACGCTCAAGTCAGAGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCTGGAAGCTCCCTCGTGCCTCTCCTGTTCCG

3095 ACCCTGCCGTTACCGGATACCTGTCCGCCTTCTCCCTTCGGGAAGCGTGGCGCTTCTCATAGCTCAGCTGTAGGTATCTCAGTTCGGTGTAGGTCG

3195 TTCGCTCCAAGCTGGGCTGTGTGCACGAACCCCCGTTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGACACGA
ApaLI (3214)

3295 CTTATCGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTCTTGAAGTGGTGGCCTAACTACGGCTAC

3395 ACTAGAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAACCACCGCTGGTA

3495 GCGGTGGTTTTTTTGTGTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTCTACGGGTCTGACGCTCAGTGAA

3595 CGAAACTCACGTTAAGGGATTTTGGTCAATGGCTAGTTAATTAACATTTAAATCAGCGGCCGCAATAAAATATCTTTATTTTATTACATCTGTGTGTTG
PacI (3630) SwaI (3639) EagI (3650) NotI (3649)

3695 GTTTTTGTGTGAATCGTAACTAACATACGCTCTCCATCAAACAACGAAACAACAACTAGCAAATAGGCTGTCCCCAGTGCAAGTGCAAGGTGC

3795 CAGAACATTTCTCTATCGAA

Blasticidin

Selection antibiotic; cell culture tested

Catalog code: ant-bl-05, ant-bl-1, ant-bl-5, ant-bl-5b

<http://www.invivogen.com/blastcidin>

For research use only

Version 20J13-MM

PRODUCT INFORMATION

Contents

Blasticidin hydrochloride is supplied as a sterile filtered solution at 10 mg/ml in HEPES buffer. It is available in 4 pack sizes:

- ant-bl-05: 5 x 1 ml (50 mg)
- ant-bl-1: 10 x 1 ml (100 mg)
- ant-bl-5: 50 x 1 ml (500 mg)
- ant-bl-5b: 1 x 50 ml (500 mg)

Storage and stability

- Blasticidin is shipped at room temperature. Upon receipt it should be stored at 4 °C or at -20 °C. Avoid repeated freeze-thaw cycles.
- The expiry date is specified on the product label.

Note: Blasticidin is stable for 2 weeks at room temperature.

QUALITY CONTROL

Each lot is thoroughly tested to ensure the absence of lot-to-lot variation.

- Purity: ≥95% (HPLC)
- Endotoxin level: < 1 EU/mg
- Physicochemical characterization (pH, appearance)
- Cell culture tested: potency validated in blasticidin-sensitive and blasticidin-resistant mammalian cell lines
- Non-cytotoxicity of trace contaminants: absence of long-term effects confirmed in blasticidin-resistant cells

BACKGROUND

Blasticidin is a selection antibiotic that acts on both eukaryotic and prokaryotic cells. It is a peptidyl nucleoside antibiotic isolated from the culture broth of *Streptomyces griseochromogenes*. It specifically inhibits protein synthesis in both prokaryotes and eukaryotes by inhibiting peptide bond formation in the ribosomal machinery. Three blasticidin resistance genes have been cloned and sequenced: an acetyl transferase gene, *bls* from a blasticidin producer strain¹, and two deaminase genes, *bsr* gene from *Bacillus cereus*², and *BSD* gene from *Aspergillus terreus*³.

Both *bsr* and *BSD* genes are used as dominant selectable markers for gene transfer experiments in mammalian and plant cells. Although blasticidin was developed as a selection agent for mammalian cells, it can also be used in *E. coli*.

GENERAL GUIDELINES

Successful transfection is influenced by many factors. The health and viability of the cell line, the quality of the nucleic acid used, the transfection reagent, the duration of transfection, and the presence or absence of serum can all play a part.

SAFETY CONSIDERATIONS

Blasticidin is a harmful compound. Refer to safety data sheet for handling instructions.

SELECTION CONDITIONS

- *Escherichia coli*

E. coli is poorly sensitive to blasticidin, but transformants resistant to blasticidin can be selected on low salt LB agar medium (pH 8) supplemented with 100 µg/ml blasticidin. High pH enhances the activity of blasticidin.

- Mammalian cells

The working concentration of blasticidin for mammalian cell lines varies from 1 to 10 µg/ml, in a few cases up to 30 µg/ml. In a starting experiment we recommend to determine optimal concentrations of antibiotic required to kill your host cell line. After treatment, cell death occurs rapidly, allowing the selection of transfected cells with plasmids carrying the *bsr* or *BSD* genes in as little as 7 days post-transfection. Suggested concentrations of blasticidin for selection in some examples of mammalian cells are listed below.

Cell line	Medium	Blasticidin conc.	Ref.
CHO (Chinese hamster ovarian cells)	DMEM	5-10 µg/ml	4, 5
HEK293 (Human embryonic kidney cells)	DMEM	5-15 µg/ml	6, 7
HeLa (Human uterine cells)	DMEM	2.5-10 µg/ml	8, 9
Neuro2a (Mouse neuroblasts)	DMEM	30 µg/ml	10
THP-1 (Human monocytes)	RMPI	10 µg/ml	11

1. Perez-Gonzalez J. et al., 1990. Cloning and characterization of the gene encoding a blasticidin S acetyltransferase from *Streptovorticillum* sp. Gene. 86:129-34.
2. Izumi M. et al., 1991. Blasticidin S-resistance gene (*bsr*): A novel selectable marker for mammalian cells. Exp.Cell Res.197:229-33.
3. Kimura M. et al., 1994. Blasticidin S deaminase gene from *Aspergillus terreus* (*BSD*): a new drug resistance gene for transfection of mammalian cells. Biochim. Biophys. Acta. 1219:653-9.
4. Dorgham K. et al., 2009. An engineered CX3CR1 antagonist endowed with anti-inflammatory activity. J Leukoc Biol. 86(4):903-11.
5. LeBon L. et al., 2014. Fringe proteins modulate Notch-ligand cis and trans interactions to specify signalling states. eLife Sci. 3:e02950.
6. Tomecki R. et al., 2014. Multiple myeloma-associated hDIS3 mutations cause perturbations in cellular RNA metabolism and suggest hDIS3 PIN domain as a potential drug target. Nucleic Acids Res. 42:1270-90.
7. Edbauer D. et al., 2004. Co-expression of nicastrin and presenilin rescues a loss of function mutant of APH-1.J Biol Chem. 279:37311-5.
8. Khandelia P. et al., 2011. Streamlined platform for short hairpin RNA interference and transgenesis in cultured mammalian cells. PNAS 108:12799-804.
9. Lee HK. et al., 2007. Application of beta-lactamase enzyme complementation to the high-throughput screening of toll-like receptor signaling inhibitors. Mol Pharmacol. 72:868-75.
10. Matsumoto G. et al., 2011. Serine 403 phosphorylation of p62/SQSTM1 regulates selective autophagic clearance of ubiquitinated proteins. Mol Cell. 44:279-89.
11. Schepetkin IA. et al., 2009. Immunomodulatory activity of oenothin B isolated from *Epilobium angustifolium*. J Immunol. 183:6754-66.

TECHNICAL SUPPORT

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RELATED PRODUCTS

Product	Description	Catalog Code
Other selection antibiotics		
G418	Selection antibiotic for the <i>neo</i> gene	ant-gn-1
Hygromycin B Gold	Selection antibiotic for the <i>hph</i> gene	ant-hg-1
Puromycin	Selection antibiotic for the <i>pac</i> gene	ant-pr-1
Zeocin™	Selection antibiotic for the <i>Sh ble</i> gene	ant-zn-1
Plasmids encoding the <i>bsr</i> gene		
pMOD2-Blast	Plasmid encoding a synthetic <i>bsr</i> gene	pmod2-blast
pSELECT-blasti-LacZ	LacZ-expression plasmid selectable with blasticidin	psetb-lacz
pSELECT-blasti-mcs	Expression plasmid selectable with blasticidin	psetb-mcs
pUNO1-bsr	Expression plasmid selectable with blasticidin	puno1-bsr

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