

pFUSE2-CLIg-rk1

Plasmid featuring the constant region of the rabbit immunoglobulin kappa 1 light chain

Catalog # pfuse2-rc1k1

For research use only

Version 20J08-MM

PRODUCT INFORMATION

Content:

- 20 µg of pFUSE2-CLIg-rk1 plasmid provided as lyophilized DNA.
- 2 x 1 ml blasticidin at 10 mg/ml

Storage and Stability:

- Product is shipped at room temperature.
- Lyophilized DNA should be stored at -20°C and is stable 3 months.
- Resuspended DNA should be stored at -20°C and is stable up to 1 year.

Quality control:

- Plasmid construct has been confirmed by restriction analysis and sequencing.
- Plasmid DNA was purified by ion exchange chromatography.

Materials required for antibody generation & isotype switching

- pFUSE2-CLIg plasmid that features the constant region of the kappa or lambda light chains. pFUSE2-CLIg plasmids are selectable with blasticidin.
- pFUSE-CH1g plasmid for the constant region of the heavy chain, this plasmid is selectable with Zeocin™ (sold separately, see RELATED PRODUCTS).

GENERAL PRODUCT USE

pFUSE-CL1g and pFUSE-CH1g plasmids are designed to change a monoclonal antibody from one isotype to another, therefore, enabling the generation of antibodies with the same antigen affinity but with different effector functions (increased or reduced ADCC and CDC). Furthermore, they can be used to produce entire IgG antibodies from Fab or scFv fragments.

pFUSE-CH1g and pFUSE2-CL1g express the constant regions of the heavy (CH) and light (CL) chains, respectively. They contain a multiple cloning site (MCS) upstream of these constant regions to enable the cloning of the variable (VH and VL) regions of a given antibody. Transfection of mammalian cell lines with the recombinant pFUSE-CH1g and pFUSE2-CL1g pair allows to generate an IgG antibody that can be purified from the supernatant using the appropriate Protein A, Protein G or Protein L affinity chromatography.

Features of pFUSE2-CL1g and pFUSE-CH1g plasmids

- **hEF1-HTLV prom** is a composite promoter comprising the Elongation Factor-1α (EF-1α) core promoter¹ and the R segment and part of the U5 sequence (R-U5') of the Human T-Cell Leukemia Virus (HTLV) Type 1 Long Terminal Repeat². The EF-1α promoter exhibits a strong activity and yields long lasting expression of a transgene *in vivo*. The R-U5' has been coupled to the EF-1α core promoter to enhance stability of RNA.
- **MCS:** The multiple cloning site contains several restriction sites that are compatible with many other enzymes, thus facilitating cloning.
- **SV40 pAn:** The Simian Virus 40 late polyadenylation signal enables efficient cleavage and polyadenylation reactions resulting in high levels of steady-state mRNA³.
- **ori:** A minimal *E. coli* origin of replication to limit vector size, but with the same activity as the longer Ori.
- **CMV enh / hFerL prom:** This composite promoter combines the human cytomegalovirus immediate-early gene 1 enhancer and the core promoter of the human ferritin light chain gene. This ubiquitous promoter drives the expression of the blasticidin-resistance gene in mammalian cells.
- **EM2KC** is a bacterial promoter that enables the constitutive expression of the antibiotic resistance gene in *E. coli*. EM2KC is located within an intron and is spliced out in mammalian cells.
- **βGlo pAn:** The human beta-globin 3'UTR and polyadenylation sequence allows efficient arrest of the transgene transcription⁴.

pFUSE2-CL1g-rk1 specific features

- **Rabbit IgKC1 (Ig Kappa 1 Light constant domain):** When cloning your VL of choice in the MCS, care must be taken to preserve the integrity of the kappa 1 light chain constant region and reading frame.
- **Bsr (blasticidin resistance gene):** Resistance to blasticidin is conferred by the bsr gene from *Bacillus cereus*. The same resistance gene confers selection in both mammalian cells and *E. coli*.

References:

1. Kim DW. *et al.* 1990. Use of the human elongation factor 1 alpha promoter as a versatile and efficient expression system. 91(2):217-23. 2. Takebe Y. *et al.* 1988. SR alpha promoter: an efficient and versatile mammalian cDNA expression system composed of the simian virus 40 early promoter and the R-U5 segment of human T-cell leukemia virus type 1 long terminal repeat. Mol Cell Biol. 8(1):466-72. 3. Carswell S. & Alwine JC. 1989. Efficiency of utilization of the simian virus 40 late polyadenylation site: effects of upstream sequences. Mol Cell Biol. 9(10):4248-58. 4. Yu J. & Russell JE. 2001. Structural and functional analysis of an mRNP complex that mediates the high stability of human beta-globin mRNA. Mol Cell Biol. 21(17):5879-88.

TECHNICAL SUPPORT

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InvivoGen USA (International): +1 (858) 457-5873
InvivoGen Europe: +33 (0) 5-62-71-69-39
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PROTOCOL

Obtaining VH and VL sequences

To obtain the cDNA sequence of the VH and VL regions from an antibody producing hybridoma, total RNA or mRNA is extracted and reverse transcribed to cDNA. PCR is performed with 5' degenerate primers to anneal to the unknown VH and VL regions and the 3' primers designed to anneal to the "known" CH and CL regions. Alternatively 5' RACE can be used. The resulting amplicons must be sequenced.

Plasmid resuspension

Quickly spin the tube containing the lyophilized plasmid to pellet the DNA. To obtain a plasmid solution at 1 $\mu\text{g}/\mu\text{l}$, resuspend the DNA in 20 μl of sterile H₂O. Store resuspended plasmid at -20°C.

Cloning into pFUSE-CHIg and pFUSE2-CLIg

Once the VH and VL sequence are known, inserts for cloning into the plasmids can be generated. In pFUSE2-CLIg-rk1, the constant region of the rabbit kappa 1 light chain is preceded by a multiple cloning site containing six unique restriction sites: AgeI, EcoRV, XhoI, NcoI, Acc65I and BamHI. We recommend using the AgeI restriction site for insertion of the 5' end of the variable region including the native signal sequence. If the immunoglobulin signal sequence is unknown, pFUSE2ss and pFUSEss plasmids containing a signal sequence should be used. In pFUSE2-CLIg-rk1, use BamHI as the 3' cloning site for the VL in order to preserve the immunoglobulin kappa 1 constant amino acid sequence.

Notes:

- When generating the insert for VH, a XhoI restriction site must be introduced at the 3' end (for pFUSE-CHIg plasmid featuring the rabbit IgG (allele 3/2) heavy chain; see RELATED PRODUCTS). There is a choice of restriction sites at the 5' end.

- The 5' end of the variable region should encompass the native ATG initiation codon and the region immediately after which corresponds to the signal sequence. For proper initiation of translation, make sure that your insert contains a Kozak translation initiation sequence upstream of the ATG initiation codon such as (G/A)NNATG.

Antibody production

Cotransfect mammalian cells, such as 293 and CHO cells, with the recombinant plasmids pFUSE2-CLIg encoding the light chain and pFUSE-CHIg encoding the heavy chain. Antibody production depends greatly on the ratio of heavy chain and light chain expression. Typically, pFUSE-CHIg to pFUSE2-CLIg ratio of 2:3 is used to cotransfect mammalian cells. Since both plasmids share the same plasmid backbone, the appropriate heavy chain to light chain ratio can be easily determined by varying the quantities of plasmids.

OR

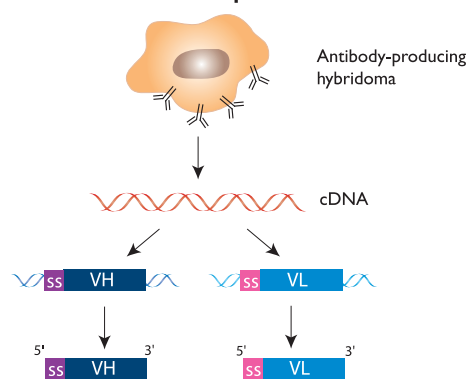
Transfect cells using a transfection agent, such as LyoVec™, with the plasmid coding for light chain and select the best clone. Following selection of the best clone, the plasmid coding for the heavy chain clone can be transfected into this clone.

Use blasticidin and Zeocin™ to select pFUSE2-CLIg and pFUSE-CHIg respectively.

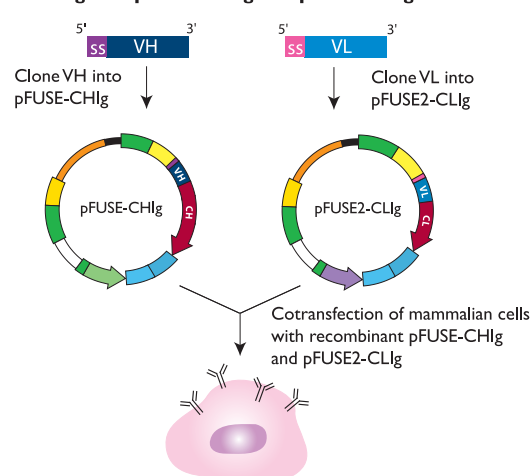
Antibody production can be analyzed by different techniques including SDS-PAGE, flow cytometry, ELISA, or a bioactivity assay.

Antibody generation using pFUSE-CHIg & pFUSE2-CLIg

1- Obtention of VH and VL sequences



2- Cloning into pFUSE-CHIg and pFUSE2-CLIg



Antibody purification

The resulting IgG antibody can be purified from the supernatant using the appropriate Protein A, Protein G or Protein L affinity chromatography.

RELATED PRODUCTS

Product	Catalog Code
pFUSE-CHIg-rG LyoVec™	pfuse-rchg lyec-12
Protein L / Agarose	gel-protl-2
Protein G / Agarose	gel-agg-5
Blasticidin	ant-bl-1

TECHNICAL SUPPORT

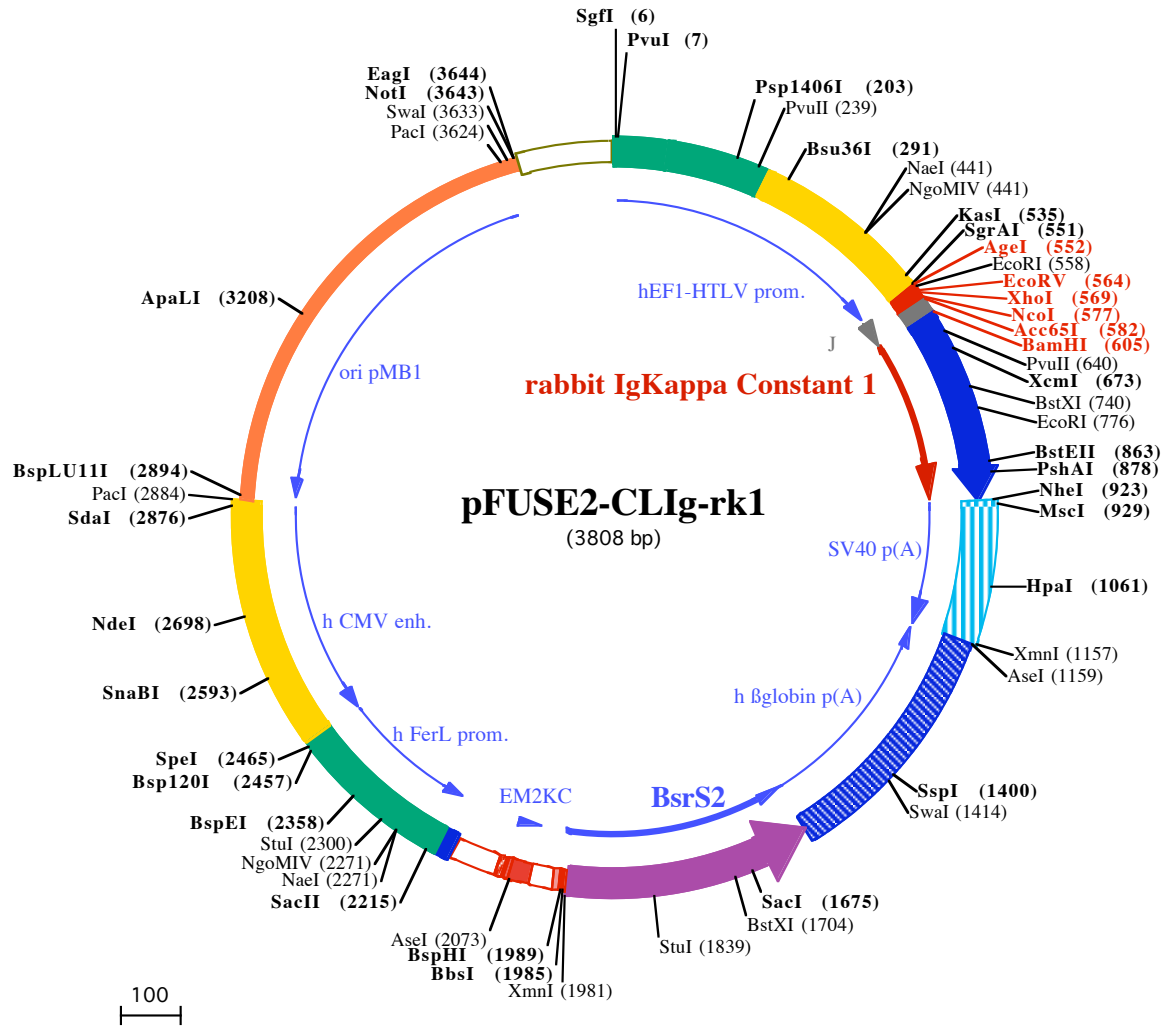
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PvuI (7)
SgfI (6)
 1 GGATCTGCGATCGCTCCGGTGCCCGTCAGTGGGCAGAGCGCACATCGCCACAGTCCCGGAGAAGTTGGGGGAGGGGTCGGCAATTGAACGGGTGCCTA
 101 GAGAAGGTGGCGCGGGTAAACTGGGAAAGTGATGTCGTGACTGGTCCGCCTTTTTCCGAGGGTGGGGGAGAACCCTATATAAGTGCAGTAGTCGCC

Psp1406I (203) **PvuII (239)** **Bsu36I (291)**
 201 GTGAACGTTCTTTTTCGCAACGGGTTTGCCGCCAGAACACAGCTGAAGCTTCGAGGGCTCGCATCTCTCCTTACGCGCCCGCCGCTACCTGAGGCC
 301 GCCATCCACGCGCGTTGAGTCGCGTTTCTGCCGCCTCCCGCCTGTGGTGCCTCCTGAAGTCGCTCCGCCGTCTAGGTAAGTTTAAAGCTCAGGTCGAGACC

NgoMIV (441)
NaeI (441)
 401 GGGCCTTTGTCCGGCGCTCCCTTGAGCCTACCTAGACTCAGCCGGCTCTCCACGCTTTGCTGACCCTGCTTGTCAACTCTACGTCTTTGTTTCGTTT

EcoRI (558)
AgeI (552) **XhoI (569)** **Acc65I (582)**
KasI (535) **SgrAI (551)** **EcoRV (564)** **NeoI (577)**
 501 TCTGTTCTGCGCCGTTACAGATCCAAGCTGTGACCGGGCGCTACCTGAGATCACCGGTGAATTCGATATCTCGAGCACCATGGGTACCAAGCTTGAAAT
 15▶ K R D P V A P T V L I F P P A A D Q V A T G T V T I V C V A N K Y
 600 CAAA**CGG**GATCCAGTTGCACCTACTGTCTCATCTTCCACCAGCTGCTGATCAGGTGGCAACTGGAACAGTCACCATCGTGTGTGGGAATAAATA
 15▶ K R D P V A P T V L I F P P A A D Q V A T G T V T I V C V A N K Y
BamHI (605) **PvuII (640)** **XcmI (673)**
 699 CTTCCCGATGTCACCGTACCTGGGAGGTGGATGGCACCACCCAAACAACCTGGCATCGAGAACAGTAAACACCGCAGAATTCGCAGATTGTACCTAC
 31▶ F P D V T V T W E V D G T T Q T T G I E N S K T P Q N S A D C T Y
BstXI (740) **EcoRI (776)**
 799 AACCTCAGCAGCACTCTGACACTGACCAGCACACAGTACAACAGCCACAAGAGTACACCTGCAAGGTGACCCAGGGCACGACCTCAGTCGTCCAGAGCT
 65▶ N L S S T L T L T S T Q Y N S H K E Y T C K V T Q G T T S V V Q S
BstEII (863) **PshAI (878)**
MscI (929)
NheI (923)
 899 TCAACAGGGGTGACTGCTAGAGCGAGCTAGCTGGCCAGACATGATAAGATACATTGATGAGTTTGGACAAACCACAACCTAGAATGCAGTAAAAAATGC
 98▶ F N R G D C •

HpaI (1061)
 999 TTTATTTGTGAAATTTGTGATGCTATTGCTTTATTTGTAACCATTATAAGCTGCAATAAACAAAGTTAACAAACAACAAATTGCATTCAATTTATGTTTC

AseI (1159)
XmnI (1157)
 1096 AGTTTCAGGGGGAGGTGTGGGAGGTTTTTAAAGCAAGTAAAACCTCTACAAATGTGGTATGGAATTAATCTCTAAAATACAGCATAGCAAACTTTAAC
 1195 CTCAAATCAAGCCTCTACTTGAATCCTTTTCTGAGGGATGAATAAGGCATAGGCATCAGGGGCTGTTGCCAATGTGCATTAGCTGTTTGCAGCCTCACC
 1295 TTCTTTTCATGGAGTTAAGATATAGTGATTTTCCCAAGGTTGAACTAGCTCTTCATTTCTTTATGTTTTAAATGCACTGACCTCCACATTCCCTTTT

SspI (1400) **SwaI (1414)**
 1395 TAGTAAAATATTCAGAAAATAATTTAAATACATCATTGCAATGAAAATAAATGTTTTTTATTAGGCAGAATCCAGATGCTCAAGGCCCTTCATAATATCCC
 1495 CCAGTTTAGTAGTTGACTTAGGGAACAAAGAACCTTTAATAGAAATTGGACAGCAAGAAAGCGAGCTTCTAGCTTTAGTTCCTGGTACTTGGAGGGG
 141▶ • N R T Y K L P
SacI (1675)
 1595 GATGAGTTCCTCAATGGTGGTTTTGACCAGCTTGCATTCTCAATGAGCACAAGCAGTCAGGAGCATAGTCAGAGATGAGCTCTCTGCACATGCCA
 133▶ I L E E I T T K V L K G N M E I L V F C D P A Y D S I L E R C M G
BstXI (1704)
 1695 CAGGGGCTGACCACCTGATGGATCTGTCCACCTCATCAGAGTAGGGGTGCCTGACAGCCACAATGGTGTCAAAGTCTTCTGCCGTTGCTCACAGCAG
 99▶ C P S V V R I S R D V E D S Y P H R V A V I T D F D K Q G N S V A S
StuI (1839)
 1795 ACCCAATGGCAATGGCTTACGACAGACAGTGACCCTGCAATGTAGGCTCAATGTGGACAGCAGAGATGATCTCCCAAGTCTTGGTCTGATGGCCG
 66▶ G I A I A E A C V T V R G I Y A E I H V A S I I E G T K T R I A A
BspHI (1989)
BbsI (1985)
XmnI (1981)
 1895 CCCGACATGGTGCTTGTGTCTCATAGAGCATGGTATCTTCTCAGTGGCGACCTCCACCAGCTCCAGATCTGCTGAGAGATGTTGAAGGCTTTCATG
 33▶ G V H H K N D E Y L M T I K E T A V E V L E L D Q Q S I N F T K M
AseI (2073)
 1995 ATGGCTCCTCctgtcaggagaggaaagagaagaaggttagtacaattgCTATAGTGAGTTGTATTATACTATGCTTATGATTAATTGTCAAATAGGG
 2093 CTGCAgggttcatagtgccacttttctgcactgccccatctctgccccctttccaggcatagacagtcagtgacttacCAAACCTCACAGGAGGGA
NgoMIV (2271)
NaeI (2271)
 2193 GAAGGCAGAAGCTTGAGACAGACCCGCGGACCCGGAAGTGCAGAGGGGACGTGGCTAGGGCGGCTTCTTTTATGGTGCAGCCGCTCGGAGGCAGGGC
SacII (2215)
StuI (2300) **BspEI (2358)**
 2293 GCTCGGGGAGGCTAGCGGCCAATCTGCGGTGGCAGGAGGCGGGGCCAAGGCCGTGCCTGACCAATCCGGAGCACATAGGAGTCTAGCCCCCGCCCC

2393 AAAGCAAGGGGAAGTCACGCGCCTGTAGCGCCAGCGTGTGTGAAATGGGGGCTTGGGGGGTTGGGGCCCTGACTAGTCAAAAACAACTCCATTGACG
Bsp120I (2457) SpeI (2465)
2493 TCAATGGGGTGGAGACTTGGAAATCCCCGTGAGTCAAACCGCTATCCACGCCATTGATGTACTGCCAAAACCGCATCATCATGGTAATAGCGATGACTA
SnaBI (2593)
2593 ATACGTAGATGTACTGCCAAGTAGGAAAGTCCCATAAAGTTCATGTACTGGGCATAATGCCAGGCGGGCCATTTACCGTCATTGACGTCAATAGGGGGCGT
NdeI (2698)
2693 ACTTGGCATATGATACACTTGTACTGCAAGTGGGCAGTTTACCCTAAATACTCCACCCATTGACGTCAATGGAAAGTCCCTATTGGCGTACTATG
Pacl (2884) SdaI (2876)
2793 GGAACATACGTCATTATTGACGTCAATGGGCGGGGTCGTTGGGCGGTGAGCCAGGCGGGCCATTTACCGTAAGTTATGTAACGCCTGCAGGTTAATTA
BspLU11I (2894)
2893 GAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAGGCGCGGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAA
2993 AATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCTGTTCCGACCC
3093 TGCCGCTTACCGGATACCTGTCCGCTTTCTCCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCC
ApaLI (3208)
3193 CTCCAAGCTGGGCTGTGTGCACGAACCCCCGTTACGCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTA
3293 TCGCCACTGGCAGCAGCCACTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGTACAGAGTTCTTGAAGTGGTGGCCTAACTACGGCTACACTA
3393 GAAGAACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAAACAAACCACCGCTGGTAGCGG
3493 TGGTTTTTTTGTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGTCTGACGCTCAGTGAACGAA
EagI (3644)
Pacl (3624) SwaI (3633) NotI (3643)
3593 AACTCACGTTAAGGGATTTTGGTCATGGCTAGTTAATTAACATTTAAATCAGCGGCCCAATAAAATATCTTTATTTTCATTACATCTGTGTGTTGGTTT
3693 TTTGTGTGAATCGTAACTAACATACGCTCTCCATCAAACAAACGAAACAAACAACTAGCAAATAGGCTGTCCCAGTGCAAGTGCAGGTGCCAGA
3793 ACATTTCTCTATCGAA

Blasticidin

Selection antibiotic; cell culture tested

Catalog code: ant-bl-05, ant-bl-1, ant-bl-5, ant-bl-5b

<http://www.invivogen.com/blastcidin>

For research use only

Version 20J13-MM

PRODUCT INFORMATION

Contents

Blasticidin hydrochloride is supplied as a sterile filtered solution at 10 mg/ml in HEPES buffer. It is available in 4 pack sizes:

- ant-bl-05: 5 x 1 ml (50 mg)
- ant-bl-1: 10 x 1 ml (100 mg)
- ant-bl-5: 50 x 1 ml (500 mg)
- ant-bl-5b: 1 x 50 ml (500 mg)

Storage and stability

- Blasticidin is shipped at room temperature. Upon receipt it should be stored at 4 °C or at -20 °C. Avoid repeated freeze-thaw cycles.
- The expiry date is specified on the product label.

Note: Blasticidin is stable for 2 weeks at room temperature.

QUALITY CONTROL

Each lot is thoroughly tested to ensure the absence of lot-to-lot variation.

- Purity: ≥95% (HPLC)
- Endotoxin level: < 1 EU/mg
- Physicochemical characterization (pH, appearance)
- Cell culture tested: potency validated in blasticidin-sensitive and blasticidin-resistant mammalian cell lines
- Non-cytotoxicity of trace contaminants: absence of long-term effects confirmed in blasticidin-resistant cells

BACKGROUND

Blasticidin is a selection antibiotic that acts on both eukaryotic and prokaryotic cells. It is a peptidyl nucleoside antibiotic isolated from the culture broth of *Streptomyces griseochromogenes*. It specifically inhibits protein synthesis in both prokaryotes and eukaryotes by inhibiting peptide bond formation in the ribosomal machinery. Three blasticidin resistance genes have been cloned and sequenced: an acetyl transferase gene, *bls* from a blasticidin producer strain¹, and two deaminase genes, *bsr* gene from *Bacillus cereus*², and *BSD* gene from *Aspergillus terreus*³.

Both *bsr* and *BSD* genes are used as dominant selectable markers for gene transfer experiments in mammalian and plant cells. Although blasticidin was developed as a selection agent for mammalian cells, it can also be used in *E. coli*.

GENERAL GUIDELINES

Successful transfection is influenced by many factors. The health and viability of the cell line, the quality of the nucleic acid used, the transfection reagent, the duration of transfection, and the presence or absence of serum can all play a part.

SAFETY CONSIDERATIONS

Blasticidin is a harmful compound. Refer to safety data sheet for handling instructions.

SELECTION CONDITIONS

- *Escherichia coli*

E. coli is poorly sensitive to blasticidin, but transformants resistant to blasticidin can be selected on low salt LB agar medium (pH 8) supplemented with 100 µg/ml blasticidin. High pH enhances the activity of blasticidin.

- Mammalian cells

The working concentration of blasticidin for mammalian cell lines varies from 1 to 10 µg/ml, in a few cases up to 30 µg/ml. In a starting experiment we recommend to determine optimal concentrations of antibiotic required to kill your host cell line. After treatment, cell death occurs rapidly, allowing the selection of transfected cells with plasmids carrying the *bsr* or *BSD* genes in as little as 7 days post-transfection. Suggested concentrations of blasticidin for selection in some examples of mammalian cells are listed below.

Cell line	Medium	Blasticidin conc.	Ref.
CHO (Chinese hamster ovarian cells)	DMEM	5-10 µg/ml	4, 5
HEK293 (Human embryonic kidney cells)	DMEM	5-15 µg/ml	6, 7
HeLa (Human uterine cells)	DMEM	2.5-10 µg/ml	8, 9
Neuro2a (Mouse neuroblasts)	DMEM	30 µg/ml	10
THP-1 (Human monocytes)	RMPI	10 µg/ml	11

1. Perez-Gonzalez J. et al., 1990. Cloning and characterization of the gene encoding a blasticidin S acetyltransferase from *Streptovorticillum* sp. Gene. 86:129-34.
2. Izumi M. et al., 1991. Blasticidin S-resistance gene (*bsr*): A novel selectable marker for mammalian cells. Exp.Cell Res.197:229-33.
3. Kimura M. et al., 1994. Blasticidin S deaminase gene from *Aspergillus terreus* (*BSD*): a new drug resistance gene for transfection of mammalian cells. Biochim. Biophys. Acta. 1219:653-9.
4. Dorgham K. et al., 2009. An engineered CX3CR1 antagonist endowed with anti-inflammatory activity. J Leukoc Biol. 86(4):903-11.
5. LeBon L. et al., 2014. Fringe proteins modulate Notch-ligand cis and trans interactions to specify signalling states. eLife Sci. 3:e02950.
6. Tomecki R. et al., 2014. Multiple myeloma-associated hDIS3 mutations cause perturbations in cellular RNA metabolism and suggest hDIS3 PIN domain as a potential drug target. Nucleic Acids Res. 42:1270-90.
7. Edbauer D. et al., 2004. Co-expression of nicastrin and presenilin rescues a loss of function mutant of APH-1. J Biol Chem. 279:37311-5.
8. Khandelia P. et al., 2011. Streamlined platform for short hairpin RNA interference and transgenesis in cultured mammalian cells. PNAS 108:12799-804.
9. Lee HK. et al., 2007. Application of beta-lactamase enzyme complementation to the high-throughput screening of toll-like receptor signaling inhibitors. Mol Pharmacol. 72:868-75.
10. Matsumoto G. et al., 2011. Serine 403 phosphorylation of p62/SQSTM1 regulates selective autophagic clearance of ubiquitinated proteins. Mol Cell. 44:279-89.
11. Schepetkin IA. et al., 2009. Immunomodulatory activity of oenothien B isolated from *Epilobium angustifolium*. J Immunol. 183:6754-66.

TECHNICAL SUPPORT

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InvivoGen USA (International): +1 (858) 457-5873

InvivoGen Europe: +33 (0) 5-62-71-69-39

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E-mail: info@invivogen.com

RELATED PRODUCTS

Product	Description	Catalog Code
Other selection antibiotics		
G418	Selection antibiotic for the <i>neo</i> gene	ant-gn-1
Hygromycin B Gold	Selection antibiotic for the <i>hph</i> gene	ant-hg-1
Puromycin	Selection antibiotic for the <i>pac</i> gene	ant-pr-1
Zeocin™	Selection antibiotic for the <i>Sh ble</i> gene	ant-zn-1
Plasmids encoding the <i>bsr</i> gene		
pMOD2-Blast	Plasmid encoding a synthetic <i>bsr</i> gene	pmod2-blast
pSELECT-blasti-LacZ	LacZ-expression plasmid selectable with blasticidin	psetb-lacz
pSELECT-blasti-mcs	Expression plasmid selectable with blasticidin	psetb-mcs
pUNO1-bsr	Expression plasmid selectable with blasticidin	puno1-bsr

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InvivoGen USA (International): +1 (858) 457-5873
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E-mail: info@invivogen.com