LFn-Rod

T3SS Inner Rod protein fused to Lethal Factor; NLRC4 inflammasome inducer

Catalog code: tlrl-rod

https://www.invivogen.com/lfn-rod

For research use only

Version 21A11-NJ

PRODUCT INFORMATION

Contents

• 50 µg of lyophilized LFn-Rod protein

• 1.5 ml endotoxin-free water

Note: B. anthracis protective antigen (PA) is not provided.

Protein construction

T3SS Inner Rod protein [M1-S101] fused to the amino-terminal domain [A34-R296] of anthrax toxin's lethal factor (LFn) protein in N-terminal.

Accession sequence: WP_000020431 (Inner Rod; PrgJ sequence)

Species: Salmonella typhimurium

Source: Sf9 insect cells

Tag: N-terminal poly-histidine (6 x His)

Total protein size: 384 a.a. (secreted form)

Molecular weight: ~ 48 KDa (SDS-PAGE)

Purification: Ni2+ affinity chromatography

Purity: >90% (SDS-PAGE)

Formulation: Lyophilized from 0.2 μ m filtered solution in 150 mM sodium chloride, 20 mM sodium phosphate buffer with 2% human serum albumin (HSA) and 5% saccharose

Storage and stability

- LFn-Rod is shipped at room temperature.
- Upon receipt, store LFn-Rod at -20°C for up to 6 months.

- Upon resuspension, store aliquots at -20°C for up to 6 months.

Avoid repeated freeze-thaw cycles.

Quality control

- Size and purity of the protein have been confirmed by SDS-PAGE.
- The biological activity has been validated using cellular assays.

• The absence of bacterial contamination (e.g. lipoproteins and endotoxins) has been confirmed using HEK-Blue[™] TLR2 and HEK-Blue[™] TLR4 cells.

PRODUCT DESCRIPTION

LFn-Rod is a model of NLRC4/NAIP inflammasome agonist¹⁻³. Inner Rod is a component of the type III secretion systems (T3SS) of intracellular bacteria described as an NLRC4/NAIP ligand¹⁻⁴. It is fused to the amino-terminal domain of *B. anthracis* lethal factor (LFn). This fusion system, when co-administred with the anthrax toxin's protective antigen (PA), allows intracellular delivery of the bacterial ligand⁵. The combination of LFn-Rod with the anthrax protective antigen (PA) is named Rod-Tox³.

METHODS

LFn-Rod resuspension (1.9 mg/ml)

 $\frac{Note:}{} Ensure you see the lyophilized pellet before resuspension. \\ - Add 26.3 \ \mu l of endotoxin-free water to the vial and gently pipette until completely resuspended.$

- Prepare aliquots and store at -20 °C.

Working concentrations: 16 ng/ml - 10 µg/ml (final concentration)

<u>Note:</u> LFn-Rod used in combination with the B. anthracis Protective antigen (PA) allows its translocation into the cytosol. The combination is sometimes referred to as Rod-Tox³.

NLRC4 INFLAMMASOME INDUCTION

LFn-Rod can be used to induce the NLRC4 inflammasome in cellular assays, such as InvivoGen's RAW-ASC and RAW-ASC KO-NLRC4 cell lines.

Production of IL-1 β by RAW-ASC-derived cells

1. Please refer to InvivoGen's technical data sheets for information regarding growth and test conditions of our cell lines.

2. Prepare test medium: DMEM without phenol red, 4.5 g/l glucose, 4 mM L-glutamine, 10% heat-inactivated FBS, 100 U/ml penicillin, 100 µg/ml streptomycin.

3. Prepare a 1.1 x 10° cells/ml suspension in test medium and add 180 μ l of cell suspension per well of a 96-well plate (~2 x 10⁵ cells/well). 4. Prime cells with 20 μ l of Pam3CSK4 (final concentration 100 ng/ml) for 3 hours at 37 °C in 5% CO₂.

5. Carefully remove medium and add 180 µl test medium.

6. Add 10 μ l of *B. anthracis* Protective antigen (PA) (final concentration 1 μ g/ml).

Note: The concentration of PA remains the same in each well.

7. Add 10 μl of LFn-Rod to the cells (final concentration 10 $\mu g/ml$ - 16 ng/ml).

8. Incubate overnight at 37 °C in 5% CO₂.

RELATED PRODUCTS

Product	Description	Cat. Code
Pam3CSK4	TLR1/2 agonist	tlrl-pms
RAW-ASC cells	Inflammasome test cells	raw-asc
RAW-ASC KO-NLRC4 <i>c</i> ells	Inflammasome test cells	raw-asc-ko-nlrc4

1. Zhao Y. et al., 2011. The NLRC4 inflammasome receptors for bacterial flagellin and type III secretion apparatus. Nature. 477(7366):596-600. 2. Kofoed E.M. & Vance R.E., 2011. Innate immune recognition of bacterial ligands by NAIPs determines inflammasome specificity. Nature. 477(7366):592-595. 3. Rauch I. et al., 2016. NAIP proteins are required for cytosolic detection of specific bacterial ligands in vivo. The Journal of Exp. Med. 213(5):657-665. 4. Worrall L.J. et al., 2011. Structural overview of the bacterial injectisome. Curr Opin Microbiology. 14(1):3-8. 5. Ballard J.D. et al., 1996. Anthrax toxin-mediated delivery of a cytotoxic T-cell epitope in vivo. PNAS. 93(22):12531-12534.

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