293/mTLR4 Cells
HEK-293 cells stably transfected with mouse TLR4 gene
Catalog # 293-mtlr4

For research use only
Version # 14H11-MM

INTRODUCTION
Toll-like receptor (TLR) 4, the first TLR identified, is the receptor for
Gram-negative lipopolysaccharide (LPS). The TLR4 gene was shown
to be mutated in C3H/HeJ and C57BL/10ScCr mice, both of which are
low responders to LPS\(^1\). However, TLR4 alone is not sufficient to
confer LPS responsiveness. TLR4 requires MD-2, a secreted molecule,
to functionally interact with LPS\(^2\).

Furthermore, a third protein, called CD14, was shown to participate in
LPS signaling, leading to NF-\(\kappa\)B translocation. This signaling is mediated
through several adaptor proteins, MyD88, TIRAP/MyD, TRIF/TICAM1
and TRAM/TICAM2\(^4\).

1. Poltorak A. \textit{et al.}, 1998. Defective LPS signaling in C3H/HeJ and C57BL/10ScCr
2. Shimazu R. \textit{et al.}, 1999. MD-2, a molecule that confers lipopolysaccharide responsiveness on Toll-
4. Fitzgerald KA. \textit{et al.}, 2003. LPS-TLR4 signaling to IRF-3/7 and NF-\(\kappa\)B

PRODUCT INFORMATION
Contents and Storage

- 1 vial of 293/mTLR4 cells (3-7 x 10^6 cells) in Freezing Medium

\textbf{IMPORTANT:} Cells are shipped frozen. If cells are not frozen upon
arrival, contact InvivoGen immediately.
- 100 µl blasticidin (10 mg/ml). Store blasticidin at 4°C for 6 months or
at -20°C for 1 year.
- 1 ml Normocin\textsuperscript{™} (50 mg/ml). Normocin\textsuperscript{™} is a formulation of three
antibiotics active against mycoplasmas, bacteria and fungi. Store
at -20°C. Product is stable for 18 months when stored at -20°C.

Handling Cells Upon Arrival
Cells must be thawed \textit{immediately} upon receipt and grown according
to handling procedures to ensure the best cell viability and assay
performance. If you are unable to thaw the cells immediately, frozen
cells may be placed in liquid nitrogen until you are ready to thaw and
propagate them, however, this may reduce cell viability.

Product Warranty
InvivoGen warrants that cells shall be viable upon shipment from
InvivoGen for a period of thirty days, provided they have been properly
stored and handled during this period.

Cell Line Stability
Cells will undergo genotypic changes resulting in reduced responsiveness
over time in normal cell culture conditions. Genetic instability is a biological
phenomenon that occurs in all stably transfected cells. Therefore, it is
critical to prepare an adequate number of frozen stocks at early passages.
293/mTLR4 cells should not be passaged more than 20 times to remain
fully efficient. 293/mTLR4 cells should be maintained in growth medium
supplemented with blasticidin. Antibiotic pressure with blasticidin is
required to maintain the plasmid coding for the mouse TLR4 gene.

Quality control
TLR4 activity is validated upon stimulation with LPS 24 hours after
cos-transfection of 293/mTLR4 cells with a mouse CD14/MD-2
expression plasmid and an NF-\(\kappa\)B-inducible SEAP (secreted embryonic
alkaline phosphatase) reporter plasmid.
These cells are guaranteed mycoplasma-free.

USE RESTRICTIONS
These cells are distributed for research purposes only.
This product is covered by a Limited Use License. By use of this product,
the buyer agrees the terms and conditions of all applicable Limited Use
Label Licenses. For non-research use, such as screening, quality control
or clinical development, contact info@invivogen.com.

293/mTLR4 Cells are designed for studying the stimulation of mouse
TLR4 (mTLR4). 293/mTLR4 cells were generated by stable transfection
of the HEK293 cell line with the mTLR4 gene. 293/mTLR4 cells do not
express the mouse MD2 and CD14 genes.

As MD-2 and CD14 are necessary for the LPS-induced responsiveness
of TLR4, the co-transfection of 293/mTLR4 cells with the MD-2 and
CD-14 is required prior to stimulation.

Notes:
- HEK293 cells express endogeneous levels of TLR1, TLR3, TLR5, TLR6
and NOD1.
- The control cell line for 293/mTLR4 cells is 293/null.

SAFETY CONSIDERATIONS
Biosafety Level: 2
293-mTLR4 cells were derived from HEK293 cells (transformed with
adenovirus 5 DNA) that require Biosafety Level 2 according to CDC
guidelines. The biosafety level may vary depending on the country.
HANDLING PROCEDURES

Required Cell Culture Media
- **Growth Medium**: DMEM (4.5 g/l glucose), 10% (v/v) fetal bovine serum (FBS), 50 U/ml penicillin, 50 µg/ml streptomycin, 100 µg/ml Normocin™, 2 mM L-glutamine
- **Freezing Medium**: DMEM (4.5 g/l glucose), 20% FBS and 10% (v/v) DMSO, 50 U/ml penicillin, 50 µg/ml streptomycin, 100 µg/ml Normocin™, 2 mM L-glutamine
- **Test Medium**: DMEM (4.5 g/l glucose), 10% (v/v) heat-inactivated FBS (30 min at 56°C), 50 U/ml penicillin, 50 µg/ml streptomycin, 100 µg/ml Normocin™, 2 mM L-glutamine

**Note**: Heat-inactivated FBS is also commercially available.

Initial Culture Procedure

The first propagation of cells should be for generating stocks for future use. This ensures the stability and performance of the cells for subsequent experiments.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol.
3. Place vials in a freezing container and store at -80°C overnight.
4. Aliquot 1 ml cells into cryogenic vials.
5. Transfer the vial contents to a T-25 tissue culture flask containing 5 ml Growth Medium.
6. Remove supernatant containing the cryoprotective agent and resuspend cells with 1 ml of Growth Medium without selective antibiotics.
7. Place the culture at 37°C in a 5% CO₂ incubator for 20-24h.

Frozen Stock Preparation

1. Resuspend cells at a density of 3-7 x 10⁶ cells/ml in Freezing Medium prepared extemporaneously with cold Growth Medium.
2. Aliquot 1 ml cells into cryogenic vials.
3. Place vials in a freezing container and store at -80°C overnight.
4. Transfer vials to liquid nitrogen for long term storage.

**Note**: If properly stored, cells should remain stable for years.

Cell maintenance

1. Maintain and subculture the cells in growth medium supplemented with 10 µg/ml blasticidin.
2. Renew growth medium 2 times a week.
3. Cells should be passaged when a 70-80% confluency is reached. Detach the cells in presence of PBS by tapping the flask or by using a cell scraper. Do not let the cell grow to 100% confluency.

**Note**: The response of 293/mTLR4 cells can be altered by the use of trypsin. Avoid trypsin to detach 293/mTLR4 cells.

DETECTION OF TLR4 STIMULATION

TLR4 stimulation can be assessed by determining the levels of IL-8, using an ELISA kit, or by measuring the activation of NF-kB. InvivoGen has developed a simple and convenient method to evaluate TLR stimulation through NF-kB activation based on the use of an NF-kB-inducible SEAP (secreted embryonic alkaline phosphatase) reporter plasmid (pNiFty-SEAP) and QUANTI-Blue™, a SEAP detection reagent. Alternatively, InvivoGen provides HEK-Blue™ mTLR4 cells (see “Related Products”), a SEAP reporter cell line expressing the mTLR4, MD-2 and CD-14 genes.

The following protocol describes a method to assess TLR4 stimulation using pNiFty-SEAP, an NF-kB-inducible SEAP reporter plasmid.

**Note**: TLR4 requires MD-2 and CD14 to signal. 293/mTLR4 cells should be co-transfected with both genes, using the pDUO2-mMD2/CD14 plasmid for example, prior to stimulation.

Day 1: Co-transfection of 293/mTLR4 cells with pNiFty-SEAP and pDUO2-mMD2/CD14 plasmids using the transfection reagent LyoVec™.
1. Prepare pNiFty-SEAP / pDUO2-mMD2/CD14 / LyoVec™ complexes by mixing 0.5 µg of each plasmid to 100 µl of LyoVec™ following the instructions provided in the technical datasheet of LyoVec™.
2. Seed 50,000 cells of per well of a flat-bottom 96-well plate in 200 µl Growth Medium.
3. Add 10 µl of pNiFty-SEAP / pDUO2-mMD2/CD14 / LyoVec™ complexes per well.
4. Incubate the plate at 37°C in a CO₂ incubator for 20-24h.

Day 2: TLR4 stimulation
1. Remove medium and replace with 180 µl of fresh Test Medium. **Note**: FBS may contain alkaline phosphatases that can interfere with SEAP quantification. The Test Medium contains heat-inactivated FBS to eliminate the activity of these thermosensitive enzymes.
2. Add 20 µl of each sample per well of a 96-well plate.
3. Add 20 µl of a positive control (e.g. LPS-EB ultrapure at 100 µg/ml) in one well.
4. Add 20 µl of a negative control (e.g. sterile endotoxin-free water) in one well.
5. Incubate the plate at 37°C in an incubator for 1-3h.

Day 3: Detection and Quantification of SEAP
1. Prepare QUANTI-Blue™ following the instructions provided in the technical datasheet.
2. Add 180 µl of QUANTI-Blue™ per well of a 96-well plate.
3. Add 20 µl of induced 293/mTLR4 cells supernatant.
4. Incubate the plate at 37°C in an incubator for 1-3h.
5. Determine SEAP levels using a spectrophotometer at 620-655 nm.

Specificity of 293/mTLR4 cells

As their parental cell line, HEK293 cells, express endogenous levels of TLR3, TLR5 and NOD1, 293/mTLR4 cells will respond to TLR3, TLR5 and NOD1 agonists, such as poly(I:C), flagellin and iE-DAP, respectively. In order to identify TLR4-specific responses, we recommend to use 293/null cells (HEK293 cells expressing a blasticidin selectable empty plasmid) as a control cell line.

RELATED PRODUCTS

<table>
<thead>
<tr>
<th>Product</th>
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<tbody>
<tr>
<td>293/null (Control cell line)</td>
<td>293-null</td>
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<tr>
<td>Blasticidin (100 mg)</td>
<td>ant-bl-1</td>
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<tr>
<td>FLA-ST Ultrapure (S. typhimurium flagellin)</td>
<td>tlr-epstfla</td>
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<td>HEK-Blue™ mTLR4 cells</td>
<td>hkb-mtlr4</td>
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<td>iE-DAP</td>
<td>tlr-dap</td>
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<td>LPS-EB Ultrapure (E. coli O111:B4 LPS)</td>
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<td>LyoVec™ (Transfection reagent)</td>
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<td>Normocin™ (Antimicrobial agent)</td>
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<td>pDUO-mMD2/CD14</td>
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<td>pNiFty-SEAP (SEAP reporter plasmid)</td>
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<td>Poly(I:C)</td>
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<td>QUANTI-Blue™ (5 pouches)</td>
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